Review

Comprehensive review of current endodontic sealers

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Endodontic sealers for non-surgical root canal treatment (NSRCT) span many compositions and attributes. This comprehensive review discusses current types of endodontic sealers by their setting reaction type, composition, and properties: zinc oxide-eugenol, salicylate, fatty acid, glass ionomer, silicone, epoxy resin, tricalcium silicate, and methacrylate resin sealers. Setting time, solubility, sealing ability, antimicrobial, biocompatibility, and cytotoxicity are all aspects key to the performance of endodontic sealers. Because sealing ability is so important to successful outcomes, the relative degree of microleakage among all the relevant sealers was calculated by way of a meta-analysis of relevant literature. Compared to AH Plus, tricalcium silicate sealers show the lowest relative microleakage among the sealers assessed, followed by silicone sealers and other non-AH Plus epoxy resin sealers. Tricalcium silicate sealers developed should ideally combine a hermetic seal with therapeutic effects.

Keywords: Endodontic sealers, Biocompatibility, Bioactivity, Dentin-sealer interface, Sealing ability

INTRODUCTION

Choosing an endodontic sealer for clinical use is a decision that contributes to the long-term success of non-surgical root canal treatment (NSRCT)¹⁾. Sealers are used as a thin tacky paste which functions as a lubricant and luting agent during obturation, allowing the core obturation material, such as gutta-percha points or other rigid materials, to slide in and become fixed in the canal^{2,3)}. Sealers can fill voids⁴⁾, lateral canals⁵⁾, and accessory canals where core obturation materials cannot infiltrate^{6,7)}. If the sealer does not perform its function, microleakage may cause NSRCT failure via clinically undetectable passage of bacteria, fluids, molecules or ions between the tooth and restorative material^{8,9)}. Knowing the qualities and characteristics of an endodontic sealer is critical to determining the best selection and application for each clinical case.

Endodontic sealers are categorized by composition based on setting reaction and composition: zinc oxideeugenol, salicylate, fatty acid, glass ionomer, silicone, epoxy resin, tricalcium silicate, and methacrylate resin sealer systems (Table 1). Some novel sealers contain fillers or ceramic powders including calcium hydroxide, mineral trioxide aggregate (MTA), and calcium phosphate; however, they are fundamentally composed of the above sealer matrices. Until recently, many review articles were published within sealer types¹⁰⁻¹⁶. However, few reviews have been published that cover all sealer $types^{17,18}$. Therefore, in this comprehensive review, a historical perspective of each sealer type will be discussed first, followed by a description of the properties of all sealer types, such as setting time and solubility, sealing ability, antimicrobial activity, and biocompatibility and cytotoxicity. Sealer attributes

such as the rheology $^{19)}$, radiopacity $^{20)}$, and tooth discoloration $^{21\cdot 23)}$ have been shown to be satisfactory and will not be discussed in detail.

CURRENT ROOT CANAL SEALERS AND HISTORY

Chelate formation

Many dental luting agents set by way of a chelation reaction, the formation of metal complexes with polydentate (usually organic) ligands²⁴⁾. Two of the most common chelates used in dentistry are eugenolates and salicylates. For eugenolates, the setting reaction starts with water that hydrolyzes the zinc oxide to form zinc hydroxide. The zinc hydroxide and eugenol chelate and solidify²⁵⁾. For salicylates, the ion is calcium, usually formulated using calcium oxide. Although uncommon, fatty acids have also been used as ligands for chelate sealers, in conjunction with zinc oxide.

1. Zinc oxide-eugenol-based sealers

The zinc oxide-eugenol (ZOE) sealer formula developed by Rickert and Dixon^{26,27)} in 1931 became Kerr sealer, and the formula developed by Grossman in 1936²⁸⁾ became Proco-Sol sealer (StarDental, Lancaster, PA, USA). The ZOE sealers have been a standard in endodontics since their development, based on their long-term success. ZOE sealers contain zinc oxide powder and eugenol liquid, an essential oil derived from cloves^{29,30)}. When mixed and placed in moist root dentin, the zinc oxide and eugenol complex to form an amorphous gel³¹⁾. Residual zinc oxide powder remains in the gel, forming a rigid matrix³²⁾. Some of these powder-liquid sealers contain silver in the powder component (Kerr formula) which has caused darkening of the teeth. Silver-free formulas that avoid staining were developed to address this issue; including

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Туре	Product name (manufacturer, country)		Composition				
	Pulp Canal Sealer	Powder	wder Zinc oxide, Precipitated silver, Oleo resin, Thymol iodide				
	(Kerr, USA)	Liquid	Oil of cloves, Canada balsam				
	Proco-Sol	Powder	Zinc oxide, Staybelite resin, Bismuth subcarbonate, Barium sulfate				
	(StarDental, USA)	Liquid	Eugenol, Sweet oil of almond				
	Tubli-Seal	Base	Zinc oxide, Bismuth trioxide, Oil+wax, Thymol iodide, Barium sulfate				
	(Kerr, USA)	Catalyst	Eugenol, Polypale resin, Annidalin				
	Endofill (Dentsply Petrópolis Ind, Bragil)	Powder	Zinc oxide, Hydrogenated resin, Bismuth subcarbonate, Barium sulfate, Sodium borate, Dexamethasone Acetate, Hydrocortisone Acetate, Polyoxymethylene, Thymol lodide				
	Brazil)	Liquid	Eugenol, Oil of sweet almonds				
	Rocanal 2 (La Maison, Switzerland)	Powder	Zinc oxide, Titanium oxide, Orthophenylphenol, Calcium tungstate				
		Liquid	Eugenol				
Zinc	Canals (Showa Yakuhin Kano, Japan)	Powder	Zinc oxide, Barium sulfate, Bismuth subcarbonate, Rosin				
oxide-eugenol		Liquid	Clove oil, Olive oil				
	Nishika Canal Sealer Eugenol (Nippon Shika Yakuhin, Japan)	Paste A	Eugenol, Rosin, Ester gum				
		Paste B	Olive oil, Zinc oxide, Bismuth subcarbonate				
	Master-Dent Root Canal Sealer (Dentonics, USA)	Powder	Zinc oxide, Staybelite resin, Bismuth subcarbonate, Barium sulfate, Sodium borate				
		Liquid	Eugenol				
	Pulpdent Root Canal	Powder	Zinc oxide, Calcium phosphate, Zinc stearate, Barium sulfate				
	Sealer (Pulpdent, USA)	Liquid	Eugenol, Canada balsam				
	CRCS (Ivoclar Vivadent, Lichtenstein)	Powder	Zinc oxide, Calcium hydroxide, Bismuth dioxide, Barium sulfate				
		Liquid	Eugenol, Eucalyptol				
	Bioseal (OGNA Pharmaceuticals, Italy)	Powder	Zinc oxide, Natural resin, Calcium hydroxide, Barium sulfate, Hydroxyapatite, Thymol iodide, Zinc acetate				
		Liquid	Purified oleoresin, Bi-distilled eugenol				
		Base	N-ethyltoluenesulfonamide, Calcium oxide, Zinc oxide, Silicon dioxide				
Solioplata	Sealapex (Kerr, USA)	Catalyst	Methyl salicylate, 2,2-dimethylpropane-1,3-diol, Isobutyl salicylate, Bismuth trioxide, Titanium dioxide pigment, Zinc stearate				
Sancylate		Paste A	Methyl salicylate, Butylene glycol, Colophony, Bismus trioxide, Fumed silica, Titanium dioxide,				
	MTA Fillapex (Angelus, Brazil)	Paste B	Fumed silica, Titanium dioxide, Tricalcium silicaate, Dicalcium silicate, Calcium oxide, Tricalcium alminate, Pentaerythritol rosinate, p-Toluenesulfonamide				

Table 1 Endodontic sealer types

Table 1	continued
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Salicylate	Apexit	Base	Calcium hydroxide, Zinc oxide, Calcium oxide, Silicon dioxi Zinc stearate, Hydrogenised colophony,Tricalcium phospha Polydimethylsiloxane				
	(Ivociar Vivadent, Lichtenstein)	Activator	Trimethylhexanediol disalicylate, Bismuth carbonate basic, Bismuth oxide, Silicon dioxide, 1,3 Butanediol disalicylate, Hydrogenised colophony, Tricalcium phosphate, Zinc stearat				
	Apexit plus	Base	Hydrated collophonium, Calcium hydroxide, Calcium oxide, Silicon dioxide, Phosphoric acid alkyl ester				
	Lichtenstein)	Activator	Disalicylate, Bismuth hydroxide, Bismuth carbonate, Silicon dioxide, Phosphoric acid alkyl ester				
Zinc oxide-fatty acid	Canals-N	Powder	Zinc oxide, Bismuth subcarbonate				
	(Showa Yakuhin Kano, Japan)	Liquid	Fatty acids, Propylene glycol				
	Nogenol (GC America, USA)	Base	Zinc oxide, Barium sulfate, Bismuth oxychloride, Vegetable oi				
		Catalyst	Lauric acid, Chlorothymol, Hydrogenated rosin, Methyl abietate, Salicylic acid				
Glass ionomer	Ketac-Endo (3M ESPE, USA)	Powder	Calcium alminium lanthanum fluorosilicate glass, Calcium volframate, Silicic acid, Pigments				
		Liquid	Water, Tartaric acid, Polyethylene polycarbonic acid/ Maleic acid, copolymer				
	GuttaFlow 2 (Coltene/Whaledent,	Base	Zirconium oxide, Polymethylvinylsiloxane, Polymethylhydrogensiloxane, Gutta-percha				
Silicono	USA)	Catalyst	Zirconium oxide, Polymethylvinylsiloxane, Platinum catalyst				
Silicone	RoekoSeal (Coltene/Whaledent, USA)	Base	Zirconium oxide, Polymethylvinylsiloxane, Polymethylhydrogensiloxane				
		Catalyst	Zirconium oxide, Polymethylvinylsiloxane, Platinum catalyst				
Epoxy resin	AH 26 (Dentsply Sirona, Germany)	Powder	Bismuth oxide, Hexamethyleneteramine, Silver powder, Titanium oxide				
		Paste	Bisphenol A diglycidyl ether				
	AH Plus (Dentsply Sirona, Germany)	Paste A	Bisphenol A epoxy resin, Zirconium oxide, Bisphenol F epoxy resin, Calcium tungstate, Iron oxide, Silica				
		Paste B	N,N-dibenzyl-5-oxanonadiamin-1,9, Amantiameamine, Tricyclodecane-diamine, Calcium tungstate, Zirconium oxide				
	Adseal (Meta Biomed, Korea)	Base	Bisphenol A diglycidyl ether –bisphenol A copolymer, 2-Hydroxyethyl salicylate, Calcium phosphate, Bismuth subcarbonate, Zirconium oxide				
		Catalyst	Poly(1,4-butanediol)bis(4-aminobenzoate), Triethanolamine, Calcium phosphate, Bismuth subcarbonate, Zirconium oxide, Calcium oxideopolymer				
	Acroseal (Septodont, France)	Base	Bisphenol A diglycidyl ether, Calcium hydroxide, Bismus subcarbonate				
		Catalyst	Hexamehtylenetetramine, Venice turpentine, Enoxolone				
	MM seal (Micro-Mega, France)	Base	Epoxy oligomer resin, Ethylene glycol salicylate, Calcium phosphate, Bismuth subcarbonate, Zirconium oxide				
		Catalyst	Poly aminobenzoate, Triethanolamine, Calcium phosphate, Bismuth subcarbonate, Zirconium oxide, Calcium oxide				

Table 1 continued

-	Grey & Neo MTA Plus (NuSmile Avalon Biomed,	Powder	Tricaclium silicate, Tantalite, Dicalcium silicate, Calcium sulfate, Silica			
	USA)	Liquid	Water-based gel			
	BioRoot RCS	Powder	Tricalcium silicate, Zirconium oxide			
	(Septodont, France)	Liquid	Aqueous solution of calcium chloride			
	Endo CPM Sealer (EGEO, Argentina)	Powder	Silicon dioxide, Calcium carbonate, Bismuth trioxide, Barium sulfate, Propylene glycol alginate, Sodium citrate, Calcium chloride			
Tricalcium		Liquid	Water-based gel			
silicate (MTA/ Bioceramic)	iRoot SP/ EndoSequence BC/ Total Fill BC/ Edge Endo Sealer (Innovative Bioceramix, Canada)	One paste	Zirconium oxide, Calcium silicates, Calcium phosphate, Calcium hydroxide, Filler, Thickening agents			
	Ceraseal (MetaBiomed, Korea)	One paste	Calcium silicates, Zirconium oxide, Thickening agent			
	Endoseal MTA (Maruchi, Korea)	One paste	Calcium silicates, Calcium aluminates, Calcium sulfate, Radiopacifier, Thickening agent			
	Bio-C Sealer (Angelus, Brazil)	One paste	Calcium silicates, Calcium aluminates, Calcium oxide, Zirconium oxide, Ferric oxide, Silicon dioxide, Thickening age			
	EndoREZ (Ultradent, USA)	Base	UDMA, Benzoyl peroxide			
		Catalyst	Triethylene glycol dimethacrylate, p-Tolyldiethanolamine			
- Methacrylate _ resin -	Epiphany (Resilon Research, USA)	Paste A	[After mixing] UDMA, PEGDMA, EBPADMA, Bis-GMA, Barium borosilicate glasses treated with silane, Barium sulfate,			
		Paste B	Silica, Calcium hydroxide, Bismuth oxychloride, Thiosinamir Cumene hydroperoxide, Photo initiator, Stabilizers, Pigment			
	MetaSEAL	Powder	Bismuth carbonate, Organic filler, Sodium sulfinate			
	(Parkell, USA)	Liquid	4-META/HEMA, Dimethacrylates, Photoinitiator, Water			
	Super-Bond RC Sealer (Accel) (Sun Medical, Japan)	Powder	Zirconiumdioxide, Poly methyl methacrylate (PMMA)			
		Liquid	Methyl methacrylate (MMA), 4-META			
		Catalyst	Tributyl borane oxide (TBB), Hexane/Ethanol			

UDMA: urethane dimethacrylate, PEGDMA: polyethyleneglycol dimethacrylate, EBPADMA: ethoxylated bisphenol A dimethacrylate, Bis-GMA: bisphenol A-glycidyl methacrylate, 4-META: 4-methacryloxyethy trimellitate anhydride, HEMA: 2-hydroxyethyl methacrylate

Wach's Paste, the Grossman formulas, Proco-Sol sealer, followed by Tubli-Seal sealer (Kerr, Orange, CA, USA). ZOE sealers remain popular because of slow set, low cost, antibacterial properties, and ease of use³³. Although Roth sealer (Roth International, Chicago, IL, USA) was discontinued in 2018, many are currently commercially available: Pulp Canal Sealer (Kerr), Proco-Sol sealer, Tubli-Seal sealer, Endofill (Dentsply Petrópolis Ind, Rio de Janeiro, Brazil), Rocanal 2 (La Maison, Balzers, Switzerland), Canals (Showa Yakuhin Kano, Tokyo, Japan), Nishika Canal Sealer Eugenol (Nippon Shika Yakuhin, Shimonoseki, Japan), Master-Dent Root Canal Sealer (Dentonics, Charlotte, NC, USA), and Pulpdent Root Canal Sealer (Pulpdent, Watertown, MA, USA).

Variations in ZOE sealers have been introduced over several decades. A ZOE-containing paraformaldehyde sealer was developed but was unsuccessful because formaldehyde causes coagulative necrosis, and residual formaldehyde disrupts local repair of affected areas³⁴; this sealer was toxic to periradicular tissues³⁵ and contraindicated. Sargenti introduced N2 sealer in 1973³⁶, which contained lead and mercury. The toxic metals were reported to be found in distant organ systems, having migrated from the radicular spaces³⁷. N2 was not cleared by the U.S. Food and Drug Administration³⁸⁾.

ZOE sealers are also a common matrix for sealers with therapeutic additives. For example, Calciobiotic Root Canal Sealer, CRCS, (Coltene/Whaledent, Cuyahoga Falls, OH, USA), is a ZOE sealer marketed as a "calcium hydroxide sealer"³⁹. Bioseal (OGNA Pharmaceuticals, Muggiò, Italy) is a ZOE-based sealer with added hydroxyapatite⁴⁰, but no special therapeutic effects have been reported.

2. Salicylate-based sealers

Salicylate-based sealers are typically referred to by their marketed therapeutic additives instead of by their composition. For example, Sealapex (Kerr) and Apexit/ Apexit Plus (Ivoclar Vivadent, Schaan, Lichtenstein) are examples of a calcium-hydroxide-containing salicylate sealers. Calcium hydroxide $[Ca(OH)_2]$ is both alkaline and antimicrobial, desirable qualities for a therapeutic sealer⁴¹⁾. However, calcium hydroxide does not set and is slightly soluble in water. It must be used within a matrix to be an effective sealer⁴²⁾. Sealers containing calcium hydroxide were intended to promote osteogenesis and cementogenesis as well as create an antimicrobial environment43). So-called "calcium hydroxide sealers" are often placed in their own sealer type categories when differentiating sealers. Despite this, all traditional "calcium hydroxide sealers" are composed of another luting matrix.

Unfortunately, Sealapex and Apexit/Apexit Plus have not demonstrated the clinical effects desired^{11,44}, while Sealapex and calcium hydroxide encourage apical closure by cementum deposition⁴⁵. The solvation of calcium hydroxide is required if therapeutic effects are to be achieved^{11,41,46,47}. Effective sealers, however, should not be soluble and should remain intact for as long as possible⁴⁸.

Similar to the above sealers, MTA Fillapex (Angelus, Londrina, Brazil) is a unique salicylate resinbased sealer that contains 15% MTA powder⁴⁹. MTA Fillapex should not be regarded as a tricalcium silicate (MTA, a bioactive ceramic) sealer since its composition is primarily resin⁴⁹. However, many researchers have wrongly referred to this sealer as "MTA-based".

3. Fatty acid-based sealers

Eugenol is known to be a cytotoxic agent that affects a cell's membrane and respiratory functions, and clinician preparation of ZOE sealers can also affect cytotoxic outcomes⁵⁰⁻⁵³⁾. As a result, non-eugenol zinc oxide sealers were developed to avoid issues with post-operative healing. Fatty acids are used instead of eugenol as chelating agents, although the structure of their metal complexes are typically less defined and consistent than with eugenolates and salicylates by nature of their mixed compositions. Canals-N (Showa Yakuhin Kako) is a fatty acid-zinc oxide sealer that uses linoleic acid, isostearic acid, and rosin²⁹⁾. Rosin contains several resin acids, the most abundant being abietic acid, which are derived from coniferous trees⁵⁴⁾. Nogenol (GC America, Alsip, IL, USA) is another fatty acid-zinc oxide sealer

made with lauric acid.

Ionomer formation

1. Glass ionomer-based sealers

Glass ionomer sealer products are made by mixing a fine silicate glass powder with polyacrylic and related acids. When mixed, they form repeating subunits of organic monomer and inorganic ions, creating an ionomer⁵⁵. These materials are used for cements and restoratives in dentistry. Glass ionomer cement sealer, KT-308 (GC, Tokyo, Japan), releases fluoride to prevent decay and bond to tooth structure⁵⁶, but this product is no longer commercially available. Ketac-Endo (3M ESPE, St. Paul, MN, USA), a glass ionomer sealer, is available in some parts of the world.

Polymer formation by addition reaction

Silicone and epoxy resin-based sealers both polymerize by way of addition reactions. Addition reactions are differentiated from other polymerization reactions because they co-generate other products (usually water)⁵⁷⁾. Silicone-based sealers form a three-dimensional polymer network by addition polymerization as a series of cross-linkage between divinylpolysiloxane and polymethylhydrosiloxane with a platinum salt as the catalyst⁵⁸⁾. Epoxy resin-based sealers follow a more traditional organic addition reaction, where epoxide monomers react with amines to create a rigid material⁵⁹⁾.

1. Silicone-based sealers

In 1972, Davis et al. used injectable silicone impression material into the prepared root canals⁶⁰⁾. Silicone-based sealers are composed of polymethyvinylsiloxane containing а platinum salt and polymethylhydrogensiloxane and set by addition reaction between vinyl groups attached to polydimethylsiloxane chain and hydrosilyl groups attached to polydimethylsiloxane chain, forming polymer⁵⁸⁾. GuttaFlow, GuttaFlow 2, and RoekoSeal (Coltene/Whaledent) are examples of silicone-based sealers^{61,62)}. GuttaFlow is triturator-mixed and requires the use of a single master cone whereas GuttaFlow 2 and RoekoSeal are auto-mix.

2. Epoxy resin-based sealers

Epoxy resin was invented in 1938 by P. Castan, a Swiss chemist of de Trey (Zurich, Switzerland), and AH 26 was developed by the same company during 1940s. A prototype AH 26 was clinically tested in the early 1950s⁶³. Guttuso studied AH 26 using rat in 1963 and found moderate tissue response in 16 days⁶⁴. Feldmann and Nyborg found AH 26, implanted after one day hardening, caused much more tissue irritation than did pure silver in rabbit study in 1964⁶⁵. In 1993, Spångberg *et al.* reported that AH 26 releases formaldehyde⁶⁶, which recommended transition from AH 26 to AH Plus, which does not release formaldehyde. Epoxy resin-based sealers, such as AH 26 and AH Plus (Dentsply Sirona, Konstanz, Germany), are composed of low molecular weight epoxy resins and amines and set by addition reaction between epoxide groups attached to epoxy resins and amines to form polymer.

AH 26 exists in a powder-paste mixture while AH Plus exists in a paste-paste mixture. When sold in an automatic mixing syringe, AH Plus is known as AH Plus Jet. In the United States, AH Plus and AH Plus Jet are sold under several other names, including ThermaSeal Plus and Ribbon sealer, respectively. AH Plus is also known as TopSeal in Europe, Central America, and South America. Adseal (Meta Biomed, Cheongju, Republic of Korea), Acroseal (Septodont, Saint-Maur-des-Fossés, France), and MM seal (Micro-Mega, Besançon, France) are also commercially available.

Hydration

1. Tricalcium silicate-based (MTA/bioceramic) sealers Introduced by Torabinejad and White in the 1990s⁶⁷, MTA is a ceramic cement based on the hydraulic powders of tricalcium silicate and dicalcium silicate. These ceramic powders are the same ceramic phases present in Portland cement^{68,69}, but the dental products are more pure, finer powders, and include radiopaque excipients. Calcium silicate cements and calcium hydroxide are bioactive; that is both ceramics release calcium and hydroxide ions47). The ions induce the formation of hydroxyapatite on their surface when body fluids (or synthetic body fluids) are present. ProRoot MTA Gray (Dentsply Sirona, Johnson City, TN, USA) was the original MTA product, marketed in since 1997, but it was only used as a root-end filling material or perforation fill, not as a sealer.

Since their introduction the tricalcium silicate-based materials have been primarily used for perforation repair, retrograde root canal filling after an apicoectomy^{70,71}, pulp capping⁷²⁾, and pulpotomies⁷²⁾. Bismuth oxide, zirconia, tantalum oxide, barium zirconate have been used for radiopacity^{73,74}). Advantages of the tricalcium silicate products include sealing by HA formation and biocompatibility75-78). When mixed with water, tri- and dicalcium silicate powders react and form a hydrated matrix with embedded calcium hydroxide. The calcium and hydroxide ions continue to be released for about one month after setting⁴⁷⁾. The high pH causes the phosphate ions in body fluids to precipitate hydroxyapatite^{79,80)} at the surface. Tricalcium silicate-based sealers have been reported to cause the deposition of apatite-like crystals in the apical and middle thirds of canal walls^{81,82)}. While ProRoot MTA is not suitable as an endodontic sealer, Grey & NeoMTA Plus (NuSmile Avalon Biomed, Houston, TX, USA) are indicated for sealing⁸³⁾. Since the MTA Plus product introduction, other powderliquid commercial tricalcium silicate sealers have been introduced: BioRoot RCS (Septodont) and Endo CPM Sealer (EGEO, Buenos Aires, Argentina).

When Grossman published his eleven criteria of an ideal root canal sealer in 1982⁴⁸, endodontic sealers always consisted of a powder and liquid, but twopaste and single component materials are currently commercially available. Single-paste tricalcium silicatebased sealers are gaining popularity in clinical practice because they are easy to use, despite their high cost. iRoot SP (Innovative Bioceramix, Vancouver, Canada), EndoSequence BC (Brasseler, Savannah, GA, USA), Total Fill BC (FKG Dentaire, La Chaux-de-Fonds, Switzerland), and Edge Endo Sealer (Edge Endo, Albuquerque, NM, USA) are the same sealer, marketed under different brand names. All four materials are from the same manufacturer (Innovative Bioceramix). The setting mechanism of single-paste tricalcium silicate-based sealers is water absorption from dentin tubules⁸⁴⁾ with the concomitant formation of HA at the surface within the canals. EndoSequence BC Sealer is used with a single-cone technique, a viable option for obturation in NSRCT^{61,85)}. Other single-paste sealers containing tricalcium silicate and organic liquids are appearing: CeraSeal (Meta Biomed), Endoseal MTA (Maruchi, Gangwon-do, Republic of Korea), and Bio-C Sealer (Angelus). Three tricalcium silicate powderliquid systems are known: NeoMTA Plus, BioRoot RCS, and Endo CPM.

Some companies have marketed the tricalcium silicate materials as "bioceramics" or "biosilicates", but these terms are too general since many dental materials are bioceramics^{15,86}. The tricalcium silicate materials are distinguished by their bioactivity; that is, their ability to form hydroxyapatite on their surface and an osteogenic effect⁸⁷.

Polymer formation by radical polymerization 1. Methacrylate resin-based sealers

The first generation of methacrylate resin-based sealers began with Hydron (Hydron Technologies, St. Petersburg, FL, USA)^{88,89}, which appeared on the market during the mid-1970s⁹⁰⁻⁹². Wichterle and Lim, contact lens researchers, developed Hydron in the 1960⁹³. It was composed of 2-hydroxyethyl methacrylate polymer gel for injection in the canal without the need for a core, such as gutta-percha. However, because of its short working time, very low radiopacity, problems associated with removal from canals, and tendency to irritate the periapical tissues, its use was discontinued in the 1980s⁸⁸.

At the beginning of the 21st century, the desire for bonding between dentin and sealing materials gave way to the second generation of methacrylate sealers. EndoREZ (Ultradent, South Jordan, UT, USA) is a dualcure sealer that does not require a dentin adhesive⁹⁴). Methacrylate resin has been used without guttapercha to create a "monoseal"; that is, a sealer which binds to radicular dentin as well as the core obturation materials⁹⁵). A monoseal is achieved when the material creates a gapless interface between the dentinal wall and rigid core (also called a monoblock)¹³).

Third-generation methacrylate-based sealers make use of formulations containing self-etching primers, beginning with Resilon/Epiphany (Resilon Research, Madison, CT, USA); functionally, this addition is analogous to the "all-in-one" adhesives used in restorative dentistry⁹⁶. The Resilon/Epiphany system was an alternative to conventional gutta-percha/sealer system^{96,97)}. The Epiphany primer etched and conditioned the dentinal surface of the canal by demineralizing it and exposing the collagen matrix⁹⁸⁾. The contents of the primer allowed the Epiphany sealer to bond covalently to the dentinal surface during polymerization. The sealer also covalently bonded to the Resilon cone, thereby achieving the monoseal desired with methacrylate-based systems. The Resilon cones contained bioactive glass, which could be resorbed. Because the dentinal wall, sealer and cone are covalently bonded, they form a single unit, known as a monoblock⁹⁹⁾. RealSeal SE (Kerr) was a commercial product similar to Epiphany^{98,100)} but with less etching ability than RealSeal¹⁰¹⁾. These systems are no longer sold because they were susceptible to degradation of

their ester bonds¹⁰²
Fourth-generation methacrylate-based sealers include a combination of self-activating etchant, primer, and sealer. Hybrid Root SEAL (Sun Medical, Shiga, Japan), also commercialized as MetaSEAL (Parkell, Edgewood, NY, USA) in the United States, is the first commercially available sealers of this generation¹⁰³. Hybrid Root SEAL hybridized dentin more resistant to low pH¹⁰⁴, which was most effective after EDTA irrigation¹⁰⁵ and may reduce microleakage¹⁰⁶.

Super-Bond RC Sealer (Accel) (Sun Medical) is a commercially available methyl methacrylate-tributyl borane (MMA-TBB) resin sealer which uses TBB as an initiator and to induce interfacial polymerization of MMA at the dentin interface^{107,108}. TBB has been shown to cause graft polymerization of MMA onto dentin collagen, creating a collagen-MMA graft polymer^{108,109}. Syudo and Hayashi in 2010 introduced a "floating with accessory point technique" using Super-Bond RC Sealer (Accel). This technique has become synonymous with the single cone technique where the master cone guttapercha point need not touch the canal walls because its "floating" in the sealer. The benefit of "floating" assures interfacial adhesion between dentin and the sealer for hermetic sealing. After placement of a floating master cone gutta-percha point and accessory points may be inserted to reduce voids/bubbles and increase interfacial contact for adhesion^{110,111}. They also noticed the mixed layer of the resin and gutta-percha at the interface of the canal walls, sealer, and gutta-percha bonded, creating monoblock.

Simultaneous treatment for root canal filling and core construction (STRC), a technique developed by Masaka *et al.*, uses MMA-TBB resin to adhere a fiber post system. The fiber post has an elastic modulus similar to dentin, unlike metal posts, making to more suitable for mimicking masticatory stress and strain¹¹². STRC uses the fiber post system replacing gutta-percha points with a minimum condensing force during the root canal obturation process. STRC is beneficial because it minimizes the number of patient clinical visits and may prevent vertical root fractures as a result of monoblock formation. An outcome study of STRC reports a five year success rate of 90.9%¹¹²). While EndoSequence BC Sealer's single-cone obturation technique in NSRCT^{61,85}) exists, STRC may prove to be a more successful concept for obturation.

PROPERTIES OF SEALERS

An ideal endodontic sealer provides a complete microscopic seal such that microbes cannot pass through the root canal system; it possesses antimicrobial activity against a range of common periodontal microbes, and it accomplishes these goals without causing an inflammatory response in host tissues or demonstrating cytotoxicity. Contemporary sealers excel for some criteria but fall short when evaluated for all of them. The ADA 57 and International Organization for Standardization (ISO) 6876 standards¹¹³⁾ provide some useful tests for measuring sealer attributes, but these tests are not sufficient to determine the performance of one sealer over another. According to the methods in the documents, antimicrobial testing is not part of these standards, and ISO 7045 is used for biocompatibility testing. Following is a proposed modified list of criteria for an endodontic sealer: 1. make a hermetic seal, 2. be tacky and preferably adhesive to dentin and obturation material between it and the canal wall when set, 3. contain fine powders, preferably for anatomical accommodation, 4. radiopacity, 5. dimensionally stable with limited changes before and after setting, 6. color stable, 7. bacteriostatic or antibacterial, 8. set slowly enough for the obturation procedure, 9. insoluble in tissue fluids, 10. biocompatible, including non-mutagenic, non-sensitizing, and noncytotoxic after setting, 11. capable of removal for retreatment by chemical or mechanical means, 12. preferably bioactive, stimulating the formation of hydroxyapatite in contact with body fluids.

Setting time and solubility

Setting time and solubility are critical components in the sealing ability of sealers. Setting time in particular is clinically important for endodontic treatment. Slow setting times allow for sealer to more readily penetrate intricate canal morphology even after treatment^{114,115)}. Faster setting times may be indicated in time sensitive situations, such as when obturation must be completed quickly or a post must be placed sooner.

Setting times for ZOE sealers have shown considerable variation. Among research studies, the setting time of Proco-Sol varies by an order of magnitude (40.5 min to 42 h). Tubli-Seal has been shown to have a setting time of approximately one hour^{19,116)}. The need for water to initiate ZOE setting may lead to variations.

ISO 6876 requires less than 3% solubility of sealers in distilled water, and ZOE sealers like Pulp Canal Sealer have met this requirement. However, for retreatment, solubility in a solvent other than water is useful. ZOE sealers showed weight losses of 5.19% in halothane, over 10 min, indicating moderate solubility in common re-treatment solvents¹¹⁷⁾.

Ketac-Endo, a glass ionomer sealer, was found to have a setting time of $2.5 h^{118}$. Glass ionomer sealers were also found to have 1.6% solubility in water, which meets

the ISO 6876 and ADA 57 limits of 3% weight loss¹¹⁹⁾. With regard to solvent solubility for re-treatment, glass ionomer sealers were the least soluble in halothane, with weight loss of less than 1% after being exposed for 10 min¹¹⁷⁾.

GuttaFlow, a silicone-based sealer, was found to have a setting time of 17.4 min, the shortest setting time of sealer types considered¹²⁰⁾. GuttaFlow had only 0.13% solubility in water, meeting the ADA and ISO specifications for solubility¹¹⁹⁾.

Setting times for tricalcium silicate-based sealers, including EndoSequence BC Sealer, also known as iRoot SP have even been shown to exceed one month¹²¹; however the setting times for BioRoot RCS, Bio-C, and CeraSeal sealers are 4, 3, and 3.5 h. Tricalcium silicate sealers such as BioRoot RCS and TotalFill BC sealer were found to have significantly higher solubility in distilled water than comparable market sealers of different compositions¹²²⁾. The solubility may be attributed to the formation of calcium hydroxide during setting of tricalcium silicates, which is dissolved in the ISO 6876 solubility test¹²³⁾. Although there are no current studies on the solubility of tricalcium silicate-based sealers in organic solvents like halothane, one study evaluating re-treatment found that the re-treatment of maxillary incisors containing EndoSequence BC Sealer with chloroform, an organic solvent that was formerly commonly used, was more facile than without¹²⁴⁾. However, the same study found EndoSequence BC Sealer had significantly more residual material remaining after retreatment compared with AH Plus¹²⁴⁾. Acids will dissolve tricalcium silicate-based sealers, but the solubility may be too slow for re-treatment. From a clinical perspective, using ultrasonic instruments is more practical than use of solvents for the tricalcium silicate sealers¹²⁵⁾.

AH 26 and AH Plus have been shown to have setting times of 34 and 8 h, respectively^{118,120}. AH Plus meets ISO solubility requirements¹²², with 0.16% solubility in water¹¹⁹. AH Plus was significantly more soluble in halothane, yielding 68% weight losses after 10 min¹¹⁷, making re-treatment viable with a solvent.

Super-Bond RC Sealer (Accel), a methacrylate resin-based sealer, was shown to have a setting time of 42 min¹²⁶⁾. Two methacrylate resin-based sealers, EndoREZ and Epiphany, were shown to have 3.5–4% solubility in water, which did not meet ADA 57 or ISO 6876 specifications¹¹⁹⁾.

Sealapex, a salicylate-based sealer, was found to have an average setting time of 58 min, which is shorter than that of ZOE sealers. Poggio *et al.* reported that Sealapex met the ISO 6876 solubility requirements¹²²⁾. Solubility in halothane for the salicylate-based sealer such as Apexit was comparable with that of ZOE sealers¹¹⁷⁾.

In summary, setting times for most sealer types were acceptable and well above one hour, with the exception of silicone-based sealers, which had markedly shorter setting times. Solubility depends on sealer matrix chemistry. For re-treatment, mechanical removal of a sealer will be useful for tricalcium silicate-based sealers and resin-based sealers.

Sealing ability

Sealing ability is of the utmost importance in sealer. Although many microleakage studies have been published, direct comparison of each sealer is difficult because experimental condition was different in each experiment/research. In many papers each sealer was tested together with AH Plus. Thus it will be convenient to compare sealing ability of each sealer by using AH Plus as a standard. The relative degree of microleakage was calculated by length of microleakage of each sealer divided by that of AH Plus. Table 2 summarizes comparisons of the degree of microleakage of different sealer types. To generate sufficient data on microleakage, an electronic search was conducted using the PubMed database (www.ncbi.nlm.nih. gov) to find studies that evaluated microleakage of the various sealers. "AH Plus", "Leakage", and "Sealing" were used as key words. Articles were limited to full-text articles written in English. The electronic search gave 152 publications. After screening by title and abstract, studies were retrieved and then read for relevance. Articles were included if they included microleakage measurements with the sealer types in question. Following discussion, 64 articles out of the 152 searched satisfied criteria and were included. Data points within the articles that compared the microleakage of different sealers were included in calculations for Table 2. Seventy two data points were used: 5 data points for tricalcium silicate (EndoSequence BC) sealers¹²⁷⁻¹³⁰, 6 data points for silicone sealers¹³¹⁻¹³⁶, 7 data points for epoxy resin sealers (other than AH Plus)^{131,137-141)}, 9 data points for salicylate sealers^{136,139,142-147}, 6 data points for zinc oxide-eugenol^{133,138,144,146,148,149)}, and 6 data points for glass ionomer (Ketac-Endo)^{132,139,142,148,150,151)}. Methacrylate resin sealers were itemized by product due to their variation: 22 data points for the Resilon/Epiphany system^{130,133,135,141,142,149,150,152-163)}, 6 data points for EndoREZ^{129,146,151,163-165)}, and 5 data points for Hybrid Root SEAL^{128,129,157)}. Some references contained data on more than one sealer; therefore, the total number of data points is more than the number of references.

Several methods have been used to assess microleakage: dye penetration, fluid filtration, glucose penetration, microbial leakage, and electrochemical leakage tests. To compare the leakage results, independent of each physicochemical method, individual measurements were converted into a ratio using AH Plus (with gutta-percha) as a standard. Sealing ability was also evaluated independent of time and as a whole. That is, time dependent measurements (e.g. microleakage at one day, one week, etc.) were averaged. For example, Bouillaguet *et al.* stated that within the 6th h of obturation, AH Plus exhibited a microleakage of 0.17 µL/min using the fluid filtration method, while GuttaFlow exhibits a microleakage of 0.08 µL/min. GuttaFlow therefore has 0.47 times the microleakage of AH Plus within the 6th h for this individual study¹³³⁾. Ratios were calculated and averaged by sealer type. Minima, maxima, and medians were determined for the

Sealers	Туре	No. of Data^ -	Degree of leakage relative to GP/AH Plus				Leakage	References	No. of
			Mean	Minimum	Maximum	Median	0		Ref
Endosequence BC, iRoot SP	Tricalcium silicate	5	0.78	0.38	1.51	0.67	Least**	127-130)	4
GuttaFlow, Roeko Seal	Silicone	6	0.83	0.19	1.84	0.65		131-136)	6
AH26, MM seal	Epoxy resin (other than AH Plus)	7	0.84	0.46	1.32	0.90		131, 137-141)	6
MTA Fillapex, Apexit, Sealapex	Salicylate	9	0.98	0.39	1.75	0.94		136, 139, 142-147)	8
Resilon/Epiphany	Methacrylate	22	0.98	0.35	2.34	0.82	Similar	130, 133, 135, 141, 142, 149, 150, 152-163)	19
Roth, Pulp Canal Sealer, ZOE	Zinc oxide- eugenol	6	1.15	0.82	1.44	1.15	to AH Plus*	133, 138, 144, 146, 148, 149)	6
Ketac-Endo	Glass ionomer	6	1.15	0.85	1.61	1.08		132, 139, 142, 148, 150, 151)	6
EndoRez	Methacrylate	6	1.17	0.70	1.58	1.18		129, 146, 151, 163-165)	6
Hybrid Root SEAL/MetaSEAL	Methacrylate	5	1.33	0.98	2.14	1.21	Most***	128, 129, 157)	3

 Table 2
 Comparisons of the degree of microleakage of different sealer types

The degree of relative microleakage of the above sealers is expressed in a ratio against AH Plus.

* AH Plus therefore has a relative degree of microleakage equal to 1.0.

** Sealers with a relative degree of microleakage less than 1.0 are considered to have less microleakage than AH Plus.

*** Sealers with a relative degree of microleakage more than 1.0 are considered to have more microleakage than AH Plus. ^ Total number of data points: 72

^^ Total number of references: 64

Since some references contain more than one sealers reported, total number of references and total number of data points are not the same.

data sets to describe the ranges of relative sealing ability in the literature. We noted that dye penetration for AH Plus/gutta-percha was: minima 0.37 mm and mean 2.49 mm in 18 tests^{131,134,136-138,140,141,146,148,151,154,158-163,165}). Dye was noted in every sealer regardless of type, suggesting that a complete microscopic seal is not achievable with contemporary sealers¹⁶⁶).

EndoSequence BC, a tricalcium silicate sealer, exhibited the lowest mean relative microleakage across the studies. In stereoscopic dye leakage tests, EndoSequence BC Sealer showed less leakage than AH Plus, Resilon/Epiphany, and ZOE-based sealers¹⁴⁶. However, dye leakage studies are inherently flawed for tricalcium silicate cements, as they absorb water until full setting. Tricalcium silicate sealers other than EndoSequence BC had mixed results. SEM studies indicated inadequate micro-sealing for Endo CPM Sealer, which had poor adaptation to canal walls¹⁶⁷. ProRoot MTA, which is not indicated as a sealer, had significantly more microleakage when used as a sealer¹⁶⁸⁾ compared with epoxy resin-based sealers AH 26 and Adseal in the dye diffusion test in extracted human teeth stereo-microscopically.

Silicone sealers, which include RoekoSeal and GuttaFlow, had the second lowest relative microleakage. These materials have a low surface tension, which allows for a high flow and low film thickness, enabling the sealer to fill intricate anatomy¹⁶⁹. RoekoSeal had the better sealing ability than GuttaFlow when measured with a dye diffusion test in extracted human teeth sectioned both horizontally and vertically¹⁶⁹. RoekoSeal has been found to expand 0.2% upon setting (exceeding the ADA 57 requirement), which may be beneficial¹⁷⁰. However, silicone sealers only seal the root mechanically (much like a polyvinylsiloxane impression material), and do not create a monoseal bond at the dentin-sealer interface.

Epoxy resin sealers other than AH Plus, namely AH 26 and MM Seal, provided nearly the same low relative

leakage as the silicone sealers; these two sealers also showed better sealing performance than AH Plus. The sealing performance of epoxy resin-based sealers can be compromised due to leaks introduced by polymerization shrinkage¹⁷¹⁾. Epoxy resin-based sealers have been shown by stereomicroscopy to have moderate sealing capacity, but superior to ZOE-based sealers¹³⁶⁾.

Salicylate resin sealers, which include MTA Fillapex, Sealapex, and Apexit, performed the closest to AH Plus. Apexit seals moderately well compared to ZOE, AH Plus, and RoekoSeal Automix, based on the cross-sectional stereomicroscopic analysis of extracted teeth¹³⁶. The salicylate-based MTA Fillapex sealer had more microleakage than conventional epoxy resin-based sealers in a dye penetration study¹⁶⁸.

ZOE sealers demonstrate more microleakage than AH Plus and any of the above-mentioned sealers. Glass ionomer sealers had an identical mean microleakage ratio to ZOE sealers¹³⁶⁾. From the maxima and minima determined for the data set, glass ionomer sealers exhibit marginally more microleakage than ZOE sealers. Glass ionomer sealers have proven to be less than satisfactory with considerable failure risk and inadequate bonding with gutta-percha^{118,136,172)}. De Gee *et al.* explained that glass ionomer sealers have low sealing capacity due to leakage pathways at the dentin-sealer interface¹¹⁸⁾.

The variations among methacrylate resin sealers is seen in Table 2. Evaluations of Resilon/Epiphany's sealing ability affirm the monoseal behind methacrylate systems. When compared with gutta-percha and other sealers in dye leakage studies, Resilon/Epiphany resulted in less microleakage up to three months after obturation¹⁷³. Bacterial leakage tests with *Streptococcus mutans* and *Enterococcus faecalis* reflect lower microleakage as well⁹⁶. The Resilon/Epiphany system performed identically to salicylate resins.

Super-Bond RC Sealer, another methacrylate resin sealer, proved to have a better microseal than both Tubli-Seal and Ketac-Endo sealers, in a dye penetration study with stereomicroscopy¹⁷⁴⁾. Resin shrinkage occurs as polymerization begins within the resin. Shrinkage of MMA-TBB resins has been shown to begin at the dentin interface, which creates superb bonding between the resin and dentin and a tight seal¹⁰⁷⁾. Interfacial initiation of the polymerization mechanism begins on the dentin side, where the resin is attracted during polymerization, and leads to the elimination of gap formation between the dentin and resin¹⁷⁵⁾. The dye penetration of Endoresin-2, an experimental MMA-TBB resin sealer, was 0.17 mm after 2 days¹⁷⁵, far less than the minimum value of 0.37 mm reported for AH Plus. Methacrylate resins have also been used with obturation material other than guttapercha to create a monoseal, with the sealer bonding to both radicular dentin and the core material⁹⁵⁾.

Antimicrobial activity

Antimicrobial activity can be directly caused by a sealer, or indirectly cause by entombing bacteria. Any endodontic sealer that does make a hermetic sealer functions to entomb bacterial within the canal and tubules, preventing communication of residual bacteria to the apical tissue¹⁷⁶⁾ However, bacteria present at the apex may not be entombed, and would be killed by an antimicrobial endodontic sealer.

Zinc oxide is a well-documented antimicrobial material because it forms a reactive oxygen species and interferes with bacterial membrane proteins¹⁷⁷⁾. ZOE sealers have better antimicrobial effects in a zone of inhibition test for Streptococcus mutans, Staphylococcus aureus, and Enterococcus faecalis, compared with multiple epoxy resin-based sealers178). Fluoride ions inhibit also bacterial growth, but glass ionomer sealers have demonstrated minimal antimicrobial activity¹⁷⁹. In general, silicone-based sealers are not antimicrobial. For instance, a zone of inhibition study with Enterococcus faecalis using of GuttaFlow 2 gave the same results as control groups without sealers¹⁸⁰. Kapralos et al. found that GuttaFlow 2 and RoekoSeal have no antibacterial activity against the planktonic growth or 24-h biofilms of Streptococcus mutans, Staphylococcus aureus, Staphylococcus epidermidis, and Enterococcus faecalis¹⁸¹⁾.

Evaluations of the antibacterial properties of MTAtype material (including non-sealer tricalcium silicates) have been confusing and sometime contradictory⁷³. In a study by Torabinejad et al., MTA was demonstrated to have an antimicrobial effect on facultative bacteria and no effect on obligate anaerobes¹⁸²⁾. Several disk diffusion studies show that MTA and Portland cement have little to no inhibitory effect on species like Staphylococcus aureus, Enterococcus faecalis, and Candida albicans¹⁸³⁻ ¹⁸⁷⁾. In disk diffusion studies, MTA and Portland cements had antimicrobial effects at least on par with other sealers like ZOE, salicylate-based (Sealapex), and epoxy resin (AH Plus)¹⁸³⁻¹⁸⁷⁾. For similar studies testing only Enterococcus faecalis, tricalcium silicate-based sealers like EndoSequence BC Sealer exhibited greater antibacterial properties than did ZOE and epoxy resin sealers¹⁸⁸⁾. It has been demonstrated that tricalcium silicate-based sealers increase the local pH through the release of calcium and hydroxide ions for adding an antimicrobial effect¹⁸⁹. In the same planktonic growth and 24-h biofilm study, Kapralos et al. also found that TotalFill BC Sealer had notable antibacterial effect on planktonic bacteria after 7 days, along with an antibacterial effect on biofilms for Staphylococcus aureus and Enterococcus faecalis¹⁸¹⁾. The same antibacterial effects of calcium hydroxide must accrue to the tricalcium silicates (MTA-type materials) because of the formation of calcium hydroxide as a reaction product from the triand di-calcium silicates.

AH Plus sealer had better antimicrobial activity only when compared with GuttaFlow but was less antimicrobial than MTA Fillapex (salicylate-based sealer) and CRCS (ZOE sealer)¹⁹⁰⁾. Compared with other sealer types, epoxy resin sealer (AH Plus) showed no significant difference in antimicrobial activity for *Enterococcus faecalis*. Zone of inhibition tests for a AH Plus were comparable with those for ZOE sealers with *Enterococcus faecalis*. However, Kapralos *et al.* found that AH Plus had the highest antibacterial activity on both planktonic and biofilm bacteria, but only lasting 24 h¹⁸¹⁾. As stated previously, tricalcium silicatebased sealers/cements (EndoSequence BC Sealer and ProRoot MTA) had higher antimicrobial activity for Enterococcus faecalis than both epoxy resin (AH Plus) and ZOE sealers^{180,188)}. The methacrylate resin-based sealer EndoREZ had the strongest antibacterial activity among comparable sealers AH Plus and Sealapex¹⁹¹⁾. In an agar diffusion test with plated strains, Micrococcus luteus (ATCC9341), Staphylococcus aureus (ATCC6538), Pseudomonas aeruginosa (ATCC27853), Candida albicans (ATCC 10231), and Enterococcus faecalis (ATCC 10541), MTA Fillapex, as well as MTA powder, were found to have antimicrobial activity against all tested strains^{185,192)}.

Despite limitations, calcium hydroxide-containing sealers have several benefits. CRCS, for example, exhibited better antimicrobial activity than epoxy resin (AH Plus) and MTA sealers when tested against *Enterococcus faecalis* on agar¹⁹⁰. Calcium-hydroxidebased sealers proved to have a greater zone of inhibition than ZOE sealers¹⁹⁰. Supercal (Ozdent, Sydney, Australia), another calcium-hydroxide-containing glycerol sealer, was more antibacterial than MTA and AH Plus sealers¹⁹³.

Biocompatibility and cytotoxicity

ZOE sealers have been shown to be both an irritant and cytotoxic agent^{194,195)} and activate a complementmediated immune response as well as significant cytotoxicity^{196,197)}. When fibroblast implanted subcutaneously in rats, eugenol inhibited the adhesion of immunocompetent cells such as macrophages and also showed more cytotoxic impact on human periodontal ligament than did ceramic powders such as zinc oxide, titanium oxide, or barium sulfate¹⁹⁸⁾, which are known to be biocompatible¹⁹⁹⁾. Because eugenol is cytotoxic and evokes an inflammatory response, zinc oxide noneugenol sealers such as Canals-N²⁹⁾ and Nogenol^{200,201)} are available in Japan and the United States.

Glass ionomer sealers have also been shown to cause inflammation when implanted subcutaneously into rats, although the inflammation was not histologically detectable after three months²⁰²⁾. Glass ionomer cement (Fuji II, GC) had lower cytotoxicity when freshly mixed compared with resin (Chem-fil II, De Trey, Wiesbaden, West Germany)²⁰³⁾. However, the same studies found that fully set, glass ionomer cement was more cytotoxic than Chem-fil II after setting, because of its fluoride ion release²⁰³⁾. Glass ionomer products have demonstrated a low level of cytotoxicity over long periods of time, indicating they consist of very biocompatible material^{204,205)}.

Silicone sealers are recognized as biocompatible, beneficial characteristic of these sealers. Significantly lower cytotoxicity was found when compared with epoxy resin sealers (AH 26 and AH Plus) during the first 11 days of fibroblast suspension cultures, and similar cytotoxicity was measured after 24 h^{206,207}. GuttaFlow was determined to be biocompatible in a fibroblast incubation test¹⁸⁰.

Tricalcium silicate-based sealers have exhibited both beneficial and deleterious effects in terms of biocompatibility¹⁶⁾. BioRoot RCS and EndoSequence BC Sealer exhibited no cytotoxic effects on human bone marrow mesenchymal cells when compared to AH Plus; EndoSequence BC Sealer has also been shown to have strong cell viability in vitro, even decreasing LPSmediated inflammation¹⁸⁸⁾. However in vivo in rats, MTA was found to be cytotoxic when histological sections of pulp tissue were examined by light microscopy at two and seven weeks²⁰⁸⁾. Another study of MTA cytotoxicity on rat subcutaneous tissue found that MTA materials had only moderate inflammation at 7 days and mild inflammation at 30 days, also suggesting that MTA induces biomineralization²⁰⁹⁾. Osteoinductive properties and cytocompatibility were superior for BioRoot RCS compared to the ZOE Kerr's Pulp Canal Sealer²¹⁰. Over time, EndoSequence BC sealer retains more pronounced cytotoxicity to osteoblast progenitors than AH Plus, even after six weeks²¹¹⁾.

Resin sealers as a whole have limited biocompatibility when unset. Unset epoxy sealers are genotoxic in mammalian cell mutation assays, attributed to residual monomer and formaldehyde²¹²⁾. Set sealers show equivocal genotoxic results, and no genotoxic activity was seen after 24 h. However, it has been noted that epoxy sealers like AH 26 release formaldehyde even two days after being mixed⁶⁶⁾. AH Plus is modified such that formaldehyde is not released³⁵⁾. High levels of inflammation have also been detected in periapical and subcutaneous tissues after the use of epoxy resin-based sealers^{198,213)}. In a rat model study, AH Plus induced milder inflammatory response than a ZOE sealer in the periapical tissue²¹⁴⁾.

Methacrylate polymer has negligible cytotoxicity when set and demonstrated cytotoxicity or inflammation only early in the setting $process^{215}$. Incompletely cured methyl methacrylate (MMA), monomer/polymer is cytotoxic although considered the least toxic monomer used in dentistry²¹⁶⁻²¹⁸. When paired with TBB, residual MMA is reduced over time^{108,215)}. Leachable materials from methacrylate-based materials, including triethyleneglycol dimethacrylate (TEGDMA), urethane dimethacrylate (UDMA), 2-hydroxyethyl methacrylate (HEMA) and polyethylene glycol dimethacrylate (PEGDMA), have shown time-dependent increases in cell death²¹⁹⁾. EndoREZ, a UDMA type of methacrylatebased material, was the most cytotoxic compared to an epoxy resin-based sealer (AH Plus) and a silicone-based sealer (RoekoSeal)²²⁰⁾. Methacrylate resin-based sealers (Real Seal and EndoREZ) have been shown to be more cytotoxic when compared with a salicylate-based (Apexit Plus) or epoxy resin-based sealer (AH Plus), based on a study testing inflammatory biomarkers²²¹⁾. However, in comparison with their epoxy resin-based counterparts (AH Plus), methacrylate-based sealers (Hybrid Root SEAL/ MetaSEAL and Super-Bond RC Sealer (Accel)) are less cytotoxic in plated cultures²²²⁾. This indicates that methyl methacrylate (MMA)-based products are more suitable in clinics than other methacrylate-based sealers.

Although their components are biocompatible, Sealapex, CRCS, and Apexit are still elicited inflammatory reactions due to poor seal²²³⁾. In vivo degradation of sealer and incomplete fills may be the reason for added inflammation in these cases²²³⁾. MTA Fillapex was also found to cause both a high level of cytotoxicity to human fibroblast cells and an increase in inflammatory mediators when freshly mixed as well as five weeks after being mixed⁷⁰⁾. Eight root canal sealers were compared for cytotoxicity for up to 72 h with human gingival fibroblasts. The tricalcium silicates and AH Plus has the highest cell viability at 24 h. However viability diminished with all after 72 h²²⁴⁾.

In summary, poor biocompatibility was noted in ZOE sealers while superior biocompatibility is an attribute for silicone-based sealers and tricalcium silicate-based sealers. AH Plus has better biocompatibility than AH 26 in epoxy resin sealers. Moderate biocompatibility is noted in methacrylate-based systems, glass ionomer sealers, and salicylate-based sealers. In epoxy resin and methacrylate resin sealers, unset sealer is less compatible than set sealer. Better biocompatibility is noted in MMA-TBB resin compared to other resin sealers.

CLINICAL IMPLICATION AND FUTURE DIRECTIONS

Pulp diagnosis as vital or necrotic is important for selection of an endodontic sealer for clinical use. In vital pulp (pulpitis) cases, the therapeutic effects of sealers are not necessary under the asepsis technique NSRCT, based on study by Kakehashi et al.²²⁵⁾. Therefore, sealers which have shown effective sealing, summarized in Table 2, are a good choice. While tricalcium silicate sealers show the least leakage, they have slow setting times. Therefore, tricalcium silicate sealers are not a good choice if post/core/build-up must occur on the same day together with endodontic obturation. In necrotic pulp cases, especially cases with large apical radiolucency, the therapeutic effects of tricalcium silicate-based sealers are useful. A medicated sealer to kill bacteria should increase the chances of long-term success. Cases of large apical radiolucency diagnosed with questionable or unfavorable prognoses are expected to benefit from sealer-driven therapeutic effects. Salicylate-based (calcium-hydroxide-containing) are good choices if post/core/build-up is performed immediately after completion of endodontic obturation. A clinician has the responsibility to decide the top priority for the patient: good sealing or a therapeutic effect.

Coronal seal by final permanent restoration is mandatory for long-term clinical success, regardless of sealer choice²²⁶⁾. The technical quality of the coronal restoration is more important than the technical quality of the endodontic treatment for apical periodontal health²²⁷⁾. Currently, manufacturers provide separate systems of endodontic obturation by gutta-percha

and sealer, post/core/build-up, and final permanent restoration. Currently, it is difficult to differentiate sealer from gutta-percha on digital radiograph system and there is a limitation of detail observation of the sealer. In the future, when the current detection level of the clinical three-dimensional cone beam computed tomography (CBCT) system (about 100 µm) approaches that of the research-grade micro computed tomography (micro-CT) machine (several $\mu m)^{228)}\!$, gutta-percha and sealer would be able to differentiate and precise observation of unfilled space or void could be possible. Sealers and obturation techniques will advance significantly together with the advancement of technology. The importance of sealers will become more of a focus in clinical treatment. Clinicians will better understand the sealer's role in preventing bacterial leakage, resulting in a successful outcome in endodontic practice.

This comprehensive review describes current types of endodontic sealers by their setting reaction type, composition, and properties. Because sealing ability is very important in achieving the best clinical outcome, the relative degree of microleakage among all the relevant sealers was calculated by way of a meta-analysis of relevant literature. Compared to AH Plus, tricalcium silicate sealers showed the lowest relative microleakage among the sealers assessed, followed by silicone sealers and other non-AH Plus epoxy resin sealers. Tricalcium silicate sealers also exhibit the most favorable antimicrobial effect and excellent biocompatibility. Future sealers developed should ideally combine a hermetic seal with therapeutic effects.

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REFERENCES

- 1) Lee M, Winkler J, Hartwell G, Stewart J, Caine R. Current trends in endodontic practice: emergency treatments and technological armamentarium. J Endod 2009; 35: 35-39.
- Caicedo R, von Fraunhofer JA. The properties of endodontic sealer cements. J Endod 1988; 14: 527-534.
- Viapiana R, Guerreiro-Tanomaru J, Tanomaru-Filho M, Camilleri J. Interface of dentine to root canal sealers. J Dent 2014; 42: 336-350.
- 4) Bodanezi A, Munhoz EA, Capelozza AL, Bernardineli N, Moraes IG, Garcia RB, *et al.* Influence of root canal sealer on the radiographic appearance of filling voids in maxillary single-rooted teeth. J Appl Oral Sci 2012; 20: 404-409.
- Almeida JF, Gomes BP, Ferraz CC, Souza-Filho FJ, Zaia AA. Filling of artificial lateral canals and microleakage and flow of five endodontic sealers. Int Endod J 2007; 40: 692-699.
- 6) Salz U, Poppe D, Sbicego S, Roulet JF. Sealing properties of a

new root canal sealer. Int Endod J 2009; 42: 1084-1089.

- 7) Jardine AP, Rosa RA, Santini MF, Wagner M, So MV, Kuga MC, *et al.* The effect of final irrigation on the penetrability of an epoxy resin-based sealer into dentinal tubules: a confocal microscopy study. Clin Oral Investig 2016; 20: 117-123.
- Coronal leakage Clinical and biological implications in endodontic success. Endodontics Colleagues for Excellence 2002: 2-7.
- Kim SY, Kim KJ, Yi YA, Seo DG. Quantitative microleakage analysis of root canal filling materials in single-rooted canals. Scanning 2015; 37: 237-245.
- Hauman CH, Love RM. Biocompatibility of dental materials used in contemporary endodontic therapy: a review. Part 2. Root-canal-filling materials. Int Endod J 2003; 36: 147-160.
- Desai S, Chandler N. Calcium hydroxide-based root canal sealers: a review. J Endod 2009; 35: 475-480.
- 12) Shrestha D, Wei X, Wu W, Ling J. Resilon: a methacrylate resin-based obturation system. J Dent Sci 2010; 5: 47-52.
- 13) Kim YK, Grandini S, Ames JM, Gu LS, Kim SK, Pashley DH, et al. Critical review on methacrylate resin-based root canal sealers. J Endod 2010; 36: 383-399.
- 14) Lotfi M, Ghasemi N, Rahimi S, Vosoughhosseini S, Saghiri MA, Shahidi A. Resilon: a comprehensive literature review. J Dent Res Dent Clin Dent Prospects 2013; 7: 119-130.
- Al-Haddad A, Che Ab Aziz ZA. Bioceramic-based root canal sealers: a review. Int J Biomater 2016; 2016: 9753210.
- 16) Primus CM, Tay FR, Niu LN. Bioactive tri/dicalcium silicate cements for treatment of pulpal and periapical tissues. Acta Biomater 2019; 96: 35-54.
- 17) Tyagi S, Mishra P, Tyagi P. Evolution of root canal sealers: An insight story. Eur J Gen Dent 2013; 2: 199-218.
- 18) Singh H, Markan S, Kaur M, Gupta G. "Endodontic Sealers": Current concepts and comparative analysis. Dent Open J 2015; 2: 32-37
- Branstetter J, von Fraunhofer JA. The physical properties and sealing action of endodontic sealer cements: a review of the literature. J Endod 1982; 8: 312-316.
- 20) Bodrumlu E, Sumer AP, Gungor K. Radiopacity of a new root canal sealer, Epiphany. Oral Surg Oral Med Oral Pathol Oral Radiol Endod 2007; 104: e59-61.
- 21) van der Burgt TP, Mullaney TP, Plasschaert AJ. Tooth discoloration induced by endodontic sealers. Oral Surg Oral Med Oral Pathol 1986; 61: 84-89.
- 22) Ioannidis K, Beltes P, Lambrianidis T, Kapagiannidis D, Karagiannis V. Crown discoloration induced by endodontic sealers: spectrophotometric measurement of Commission International de l'Eclairage's L*, a*, b* chromatic parameters. Oper Dent 2013; 38: E1-12.
- Tour Savadkouhi S, Fazlyab M. Discoloration potential of endodontic sealers: a brief review. Iran Endod J 2016; 11: 250-254.
- 24) IUPAC. Chelation. 2nd ed. McNaught A, Wilkinson A, editors. Oxford: Blackwell Scientific Publishers; 1997.
- 25) Primus CM, Shen C. Impression Materials. In: Anusavice K, Phillips R, Shen C, Rawls H, editors. Phillips' science of dental materials. 12th ed. St. Louis, Mo: Elsevier/Saunders; 2013. p. 307-339.
- 26) Rickert U, Dixon C. The controlling of root surgery. Congres Dentaire International 1931: 15-22.
- 27) Rickert U, Dixon C. The control of root surgery. Transactions of the 8th International Dental Congress 1933; 9: 1458.
- 28) Grossman LI. Filling root canals with silver points. Dental Cosmos 1936; 78: 679-687.
- 29) Araki K, Suda H, Spangberg LS. Indirect longitudinal cytotoxicity of root canal sealers on L929 cells and human periodontal ligament fibroblasts. J Endod 1994; 20: 67-70.
- 30) Fujisawa S, Murakami Y. Eugenol and its role in chronic diseases. In: Gupta S PSAB, editor. Drug Discovery from Mother Nature Advances in Experimental Medicine and

Biology. 929. Switzerland: Springer International Publishing; 2016. p. 45-66.

- Wilson AD, Mesley RJ. Zinc oxide-eugenol cements. 3. Infrared spectroscopic studies. J Dent Res 1972; 51: 1581-1588.
- 32) Wilson AD, Mesley RJ. Chemical nature of cementing matrixes of cements formed from zinc oxide and 2-ethoxybenzoic acideugenol liquids. J Dent Res 1974; 53: 146.
- 33) Civjan S, Brauer GM. Physical properties of cements, based on zinc oxide, hydrogenated rosin, o-ethoxybenzoic acid, and eugenol. J Dent Res 1964; 43: 281-299.
- 34) Spangberg L, Langeland K. Biologic effects of dental materials. 1. Toxicity of root canal filling materials on HeLa cells in vitro. Oral Surg Oral Med Oral Pathol 1973; 35: 402-414.
- 35) Leonardo MR, Bezerra da Silva LA, Filho MT, Santana da Silva R. Release of formaldehyde by 4 endodontic sealers. Oral Surg Oral Med Oral Pathol Oral Radiol Endod 1999; 88: 221-225.
- 36) Sargenti A. Debate on N2: Is N2 an acceptable method of treatment? Trans Int Conf Endod 1973; 5: 176-195.
- 37) Oswald RJ, Cohn SA. Systemic distribution of lead from root canal fillings. J Endod 1975; 1: 59-63.
- 38) FDA explains status of N2 material. J Am Dent Assoc 1992; 123: 236-237.
- 39) Cohen T, Gutmann JL, Wagner M. An assessment in vitro of the sealing properties of calciobiotic root canal sealer. Int Endod J 1985; 18: 172-178.
- 40) Gambarini G, Tagger M. Sealing ability of a new hydroxyapatite-containing endodontic sealer using lateral condensation and thermatic compaction of gutta-percha, in vitro. J Endod 1996; 22: 165-167.
- 41) Staehle HJ, Spiess V, Heinecke A, Muller HP. Effect of root canal filling materials containing calcium hydroxide on the alkalinity of root dentin. Endod Dent Traumatol 1995; 11: 163-168.
- 42) Cox CF, Suzuki S. Re-evaluating pulp protection: calcium hydroxide liners vs. cohesive hybridization. J Am Dent Assoc 1994; 125: 823-831.
- 43) Paula-Silva FW, Ghosh A, Arzate H, Kapila S, da Silva LA, Kapila YL. Calcium hydroxide promotes cementogenesis and induces cementoblastic differentiation of mesenchymal periodontal ligament cells in a CEMP1- and ERK-dependent manner. Calcif Tissue Int 2010; 87: 144-157.
- 44) Mohammadi Z, Dummer PM. Properties and applications of calcium hydroxide in endodontics and dental traumatology. Int Endod J 2011; 44: 697-730.
- 45) Holland R, Souza Vd. Ability of a new calcium hydroxide root canal filling material to induce hard tissue formation. J Endod 1985; 11: 535-543.
- 46) Ersahan S, Aydin C. Solubility and apical sealing characteristics of a new calcium silicate-based root canal sealer in comparison to calcium hydroxide-, methacrylate resin- and epoxy resin-based sealers. Acta Odontol Scand 2013; 71: 857-862.
- 47) Gandolfi MG, Siboni F, Botero T, Bossu M, Riccitiello F, Prati C. Calcium silicate and calcium hydroxide materials for pulp capping: biointeractivity, porosity, solubility and bioactivity of current formulations. J Appl Biomater Funct Mater 2015; 13: 43-60.
- 48) Grossman L. Obturation of root canal. Endodontic Practice 10th ed. Philadelphia, PA: Lea and Febiger; 1982. p. 297.
- 49) Jafari F, Jafari S. Composition and physicochemical properties of calcium silicate based sealers: A review article. J Clin Exp Dent 2017; 9: e1249-e1255.
- 50) Lindqvist L, Otteskog P. Eugenol: liberation from dental materials and effect on human diploid fibroblast cells. Scand J Dent Res 1980; 88: 552-556.
- 51) Spangberg LS. In vitro assessment of the toxicity of endodontic

materials. Int Endod J 1981; 14: 27-33.

- 52) Hume WR. Effect of eugenol on respiration and division in human pulp, mouse fibroblasts, and liver cells in vitro. J Dent Res 1984; 63: 1262-1265.
- 53) Spangberg L, Pascon EA. The importance of material preparation for the expression of cytotoxicity during in vitro evaluation of biomaterials. J Endod 1988; 14: 247-250.
- 54) Fiebach K, Grimm D. Resins, Natural. Ullmann's Encyclopedia of Industrial Chemistry (Ed)2000.
- 55) Berg JH, Croll TP. Glass ionomer restorative cement systems: an update. Pediatr Dent 2015; 37: 116-124.
- 56) Ogasawara T, Yoshimine Y, Yamamoto M, Akamine A. Biocompatibility of an experimental glass-ionomer cement sealer in rat mandibular bone. Oral Surg Oral Med Oral Pathol Oral Radiol Endod 2003; 96: 458-465.
- 57) Penczek S, Moad G. Glossary of terms related to kinetics, thermodynamics, and mechanisms of polymerization (IUPAC Recommendations 2008). Pure Appl Chem 2008; 80: 2163-2193.
- 58) Shen C. Impression Materials. In: Anusavice K, Phillips R, Shen C, Rawls H, editors. Phillips' science of dental materials. 12th ed. St. Louis, Mo: Elsevier/Saunders; 2013. p. 151-181.
- 59) Saeb MR, Bakhshandeh E, Khonakdar HA, #xe4, der E, Scheffler C, Heinrich G. Cure Kinetics of Epoxy Nanocomposites Affected by MWCNTs Functionalization: A Review. The Scientific World Journal 2013; 2013: 14.
- 60) Davis SR, Brayton SM, Goldman M. The morphology of the prepared root canal: a study utilizing injectable silicone. Oral Surg Oral Med Oral Pathol 1972; 34: 642-648.
- 61) Zhou HM, Shen Y, Zheng W, Li L, Zheng YF, Haapasalo M. Physical properties of 5 root canal sealers. J Endod 2013; 39: 1281-1286.
- 62) Gandolfi MG, Siboni F, Prati C. Properties of a novel polysiloxane-guttapercha calcium silicate-bioglass-containing root canal sealer. Dent Mater 2016; 32: e113-126.
- 63) Schroeder A. Mitteilungen über die Abschlussdichtigkeit von Wurzelfüllmaterialien und erster Hinweis auf ein neuartiges Wurzelfüllmittel. Schweizerische Monatszeitschrift fur Zahnheilkunde 1954; 64: 921-931.
- 64) Guttuso J. Histopathologic study of rat connective tissue responses to endodontic materials. Oral Surg Oral Med Oral Pathol 1963; 16: 713-727.
- 65) Feldmann G, Nyborg H. Tissue reactions to root filling materials. II. A comparison of implants of silver and root filling material AH 26 in rabiits' jaws. Odontol Revy 1964; 15: 33-40.
- 66) Spångberg LS, Barbosa SV, Lavigne GD. AH 26 releases formaldehyde. J Endod 1993; 19: 596-598.
- 67) Torabinejad M, White DJ, inventors. Tooth filling material and use. United States Patent & Trademark Office. Patent Number 5,769,638, May 16, 1995.
- 68) Torabinejad M, Watson TF, Pitt Ford TR. Sealing ability of a mineral trioxide aggregate when used as a root end filling material. J Endod 1993; 19: 591-595.
- 69) Torabinejad M, White DJ, inventors. Tooth filling material and method of use. United States patent 5,415,547. 1995.
- 70) von Arx T, Hanni S, Jensen SS. 5-year results comparing mineral trioxide aggregate and adhesive resin composite for root-end sealing in apical surgery. J Endod 2014; 40: 1077-1081.
- 71) Soundappan S, Sundaramurthy JL, Raghu S, Natanasabapathy V. Biodentine versus mineral trioxide aggregate versus intermediate restorative material for retrograde root end filling: an in vitro study. J Dent (Tehran) 2014; 11: 143-149.
- 72) Swarup SJ, Rao A, Boaz K, Srikant N, Shenoy R. Pulpal response to nano hydroxyapatite, mineral trioxide aggregate and calcium hydroxide when used as a direct pulp capping agent: an in vivo study. J Clin Pediatr Dent 2014; 38: 201-

206.

- 73) Parirokh M, Torabinejad M. Mineral trioxide aggregate: a comprehensive literature review: Part I: chemical, physical, and antibacterial properties. J Endod 2010; 36: 16-27.
- 74) Tawil PZ, Duggan DJ, Galicia JC. Mineral trioxide aggregate (MTA): its history, composition, and clinical applications. Compend Contin Educ Dent 2015; 36: 247-252; quiz 254, 264.
- 75) Roberts HW, Toth JM, Berzins DW, Charlton DG. Mineral trioxide aggregate material use in endodontic treatment: a review of the literature. Dent Mater 2008; 24: 149-164.
- 76) Parirokh M, Torabinejad M. Mineral trioxide aggregate: a comprehensive literature review: Part III: Clinical applications, drawbacks, and mechanism of action. J Endod 2010; 36: 400-413.
- 77) Torabinejad M, Parirokh M. Mineral trioxide aggregate: a comprehensive literature review--part II: leakage and biocompatibility investigations. J Endod 2010; 36: 190-202.
- 78) Darvell BW, Wu RC. "MTA"-an hydraulic silicate cement: review update and setting reaction. Dent Mater 2011; 27: 407-422.
- 79) Aguilar P, Linsuwanont P. Vital pulp therapy in vital permanent teeth with cariously exposed pulp: a systematic review. J Endod 2011; 37: 581-587.
- 80) Komabayashi T, Zhu Q, Eberhart R, Imai Y. Current status of direct pulp-capping materials for permanent teeth. Dent Mater J 2016; 35: 1-12.
- 81) Camilleri J, Montesin FE, Brady K, Sweeney R, Curtis RV, Ford TR. The constitution of mineral trioxide aggregate. Dent Mater 2005; 21: 297-303.
- 82) Gomes-Filho JE, Watanabe S, Bernabe PF, de Moraes Costa MT. A mineral trioxide aggregate sealer stimulated mineralization. J Endod 2009; 35: 256-260.
- 83) Siboni F, Taddei P, Prati C, Gandolfi MG. Properties of NeoMTA Plus and MTA Plus cements for endodontics. Int Endod J 2017; 50: e83-e94.
- 84) Koch KA, Brave DG, Nasseh AA. Bioceramic technology: closing the endo-restorative circle, Part I. Dent Today 2010; 29: 100-105.
- 85) Chybowski EA, Glickman GN, Patel Y, Fleury A, Solomon E, He J. Clinical outcome of non-surgical root canal treatment using a single-cone technique with endosequence bioceramic sealer: a retrospective analysis. J Endod 2018; 44: 941-945.
- 86) Jitaru S, Hodisan I, Timis L, Lucian A, Bud M. The use of bioceramics in endodontics — literature review. Clujul Med 2016; 89: 470-473.
- 87) Faraco IM, Jr., Holland R. Response of the pulp of dogs to capping with mineral trioxide aggregate or a calcium hydroxide cement. Dent Traumatol 2001; 17: 163-166.
- 88) Murrin JR, Reader A, Foreman DW, Beck FM, Meyers WJ. Hydron versus gutta-percha and sealer: A study of endodontic leakage using the scanning electron microscope and energydispersive analysis. J Endod 1985; 11: 101-109.
- 89) Babb BR, Loushine RJ, Bryan TE, Ames JM, Causey MS, Kim J, et al. Bonding of self-adhesive (self-etching) root canal sealers to radicular dentin. J Endod 2009; 35: 578-582.
- 90) Rising DW, Goldman M, Brayton SM. Histologic appraisal of 3 experimental root canal filling materials. J Endod 1975; 1: 172-177.
- 91) Benkel BH, Rising DW, Goldman LB, Rosen H, Goldman M, Kronman JH. Use of a hydrophilic plastic as a root canal filling material. J Endod 1976; 2: 196-202.
- 92) Kronman JH, Goldman M, Goldman LB, Coleman E, Kliment CK. Microbiologic evaluation of poly-HEMA root canal filling material. Oral Surg Oral Med Oral Pathol 1979; 48: 175-177.
- 93) Wichterle O, Lím D. Hydrophilic gels for biological use. Nature 1960; 185: 117-118.
- 94) De Munck J, Vargas M, Van Landuyt K, Hikita K, Lambrechts P, Van Meerbeek B. Bonding of an auto-adhesive luting

material to enamel and dentin. Dent Mater 2004; 20: 963-971.

- 95) Shrestha D, Wei X, Wu WC, Ling JQ. Resilon: a methacrylate resin-based obturation system. J Dent Sci 2010; 5: 47-52.
- 96) Shipper G, Orstavik D, Teixeira FB, Trope M. An evaluation of microbial leakage in roots filled with a thermoplastic synthetic polymer-based root canal filling material (Resilon). J Endod 2004; 30: 342-347.
- 97) Shipper G, Teixeira FB, Arnold RR, Trope M. Periapical inflammation after coronal microbial inoculation of dog roots filled with gutta-percha or resilon. J Endod 2005; 31: 91-96.
- 98) Teixeira FB, Teixeira EC, Thompson J, Leinfelder KF, Trope M. Dentinal bonding reaches the root canal system. J Esthet Restor Dent 2004; 16: 348-354.
- 99) Teixeira FB, Teixeira EC, Thompson JY, Trope M. Fracture resistance of roots endodontically treated with a new resin filling material. J Am Dent Assoc 2004; 135: 646-652.
- 100) Stelzer R, Schaller HG, Gernhardt CR. Push-out bond strength of RealSeal SE and AH Plus after using different irrigation solutions. J Endod 2014; 40: 1654-1657.
- 101) Kim YK, Mai S, Haycock JR, Kim SK, Loushine RJ, Pashley DH, et al. The self-etching potential of RealSeal versus RealSeal SE. J Endod 2009; 35: 1264-1269.
- 102) Payne LA, Tawil PZ, Phillips C, Fouad AF. Resilon: assessment of degraded filling material in nonhealed cases. J Endod 2019; 45: 691-695.
- 103) Lawson MS, Loushine B, Mai S, Weller RN, Pashley DH, Tay FR, et al. Resistance of a 4-META-containing, methacrylatebased sealer to dislocation in root canals. J Endod 2008; 34: 833-837.
- 104) Marin-BauzaGA, Rached-Junior FJ, Souza-Gabriel AE, Sousa-Neto MD, Miranda CE, Silva-Sousa YT. Physicochemical properties of methacrylate resin-based root canal sealers. J Endod 2010; 36: 1531-1536.
- 105) Pinna L, Loushine RJ, Bishop FD, Jr., Cotti E, Weller RN, Pashley DH, et al. Hybrid Root SEAL (MetaSEAL) creates hybrid layers in radicular dentin only when EDTA is used as the final rinse. Am J Dent 2009; 22: 299-303.
- 106) Ari H, Belli S, Gunes B. Sealing ability of Hybrid Root SEAL (MetaSEAL) in conjunction with different obturation techniques. Oral Surg Oral Med Oral Pathol Oral Radiol Endod 2010; 109: e113-116.
- 107) Imai Y, Kadoma Y, Kojima K, Akimoto T, Ikakura K, Ohta T. Importance of polymerization initiator systems and interfacial initiation of polymerization in adhesive bonding of resin to dentin. J Dent Res 1991; 70: 1088-1091.
- 108) Taira Y, Imai Y. Review of methyl methacrylate (MMA)/ tributylborane (TBB)-initiated resin adhesive to dentin. Dent Mater J 2014; 33: 291-304.
- 109) Masuhara E. [On the chemistry of a new adhesive plastic filling material]. Dtsch Zahnarztl Z 1969; 24: 620-628.
- 110) Syudo M, Hayashi Y. Proposal of the new obturation method for good sealing ability using methyl methacrylate resinbased adhesive sealer. J Jpn Dent Orient Med 2010; 29: 28-34.
- 111) Syudo M, Hayashi Y. Floating technique in root canal oburation. Nihon Shika Hyoron 2011; 71: 119-127.
- 112) Masaka N, Sekiya W, Yineda S, Masaka K, Fukushima Y, Okada T. Clinical result of the method to perform root canal filling and core construction simultaneously. Adhes Dent 2015; 33: 37-43.
- 113) ISO, 6876. Dentistry-Root canal sealing materials. Geneva: International Organization for Standardization; 2012 (E).
- 114) Allan NA, Walton RE, Schaffer M. Setting times for endodontic sealers under clinical usage and in vitro conditions. J Endod 2001; 27: 421-423.
- 115) Nunes VH, Silva RG, Alfredo E, Sousa-Neto MD, Silva-Sousa YTC. Adhesion of Epiphany and AH Plus sealers to human root dentin treated with different solutions. Braz Dent J

2008; 19: 46-50.

- 116) Grossman LI. Physical properties of root canal cements. J Endod 1976; 2: 166-175.
- 117) Whitworth JM, Boursin EM. Dissolution of root canal sealer cements in volatile solvents. Int Endod J 2000; 33: 19-24.
- 118) De Gee AJ, Wu MK, Wesselink PR. Sealing properties of Ketac-Endo glass ionomer cement and AH26 root canal sealers. Int Endod J 1994; 27: 239-244.
- 119) Donnelly A, Sword J, Nishitani Y, Yoshiyama M, Agee K, Tay FR, *et al.* Water sorption and solubility of methacrylate resinbased root canal sealers. J Endod 2007; 33: 990-994.
- 120) Camargo RV, Silva-Sousa YTC, Rosa R, Mazzi-Chaves JF, Lopes FC, Steier L, *et al.* Evaluation of the physicochemical properties of silicone- and epoxy resin-based root canal sealers. Braz Oral Res 2017; 31: e72.
- 121) Lee JK, Kwak SW, Ha JH, Lee W, Kim HC. Physicochemical properties of epoxy resin-based and bioceramic-based root canal sealers. Bioinorg Chem Appl 2017; 2017: 2582849.
- 122) Poggio C, Arciola CR, Dagna A, Colombo M, Bianchi S, Visai L. Solubility of root canal sealers: a comparative study. Int J Artif Organs 2010; 33: 676-681.
- 123) Huffman BP, Mai S, Pinna L, Weller RN, Primus CM, Gutmann JL, et al. Dislocation resistance of ProRoot Endo Sealer, a calcium silicate-based root canal sealer, from radicular dentine. Int Endod J 2009; 42: 34-46.
- 124) Oltra E, Cox TC, LaCourse MR, Johnson JD, Paranjpe A. Retreatability of two endodontic sealers, EndoSequence BC Sealer and AH Plus: a micro-computed tomographic comparison. Restor Dent Endod 2017; 42: 19-26.
- 125) Neelakantan P, Grotra D, Sharma S. Retreatability of 2 mineral trioxide aggregate-based root canal sealers: a conebeam computed tomography analysis. J Endod 2013; 39: 893-896.
- 126) Garza EG, Wadajkar A, Ahn C, Zhu Q, Opperman LA, Bellinger LL, et al. Erratum to "Cytotoxicity evaluation of methacrylate-based resins for clinical endodontics in vitro. J Oral Sci 2012; 54: 363.
- 127) Zhang W, Li Z, Peng B. Assessment of a new root canal sealer's apical sealing ability. Oral Surg Oral Med Oral Pathol Oral Radiol Endod 2009; 107: e79-82.
- 128) Onay EO, Orucoglu H, Kiremitci A, Korkmaz Y, Berk G. Effect of Er,Cr:YSGG laser irradiation on the apical sealing ability of AH Plus/gutta-percha and Hybrid Root Seal/Resilon combinations. Oral Surg Oral Med Oral Pathol Oral Radiol Endod 2010; 110: 657-664.
- 129) Ulusoy OI, Nayir Y, Celik K, Yaman SD. Apical microleakage of different root canal sealers after use of maleic acid and EDTA as final irrigants. Braz Oral Res 2014; 28.
- 130) Hegde V, Arora S. Sealing ability of three hydrophilic singlecone obturation systems: An in vitro glucose leakage study. Contemp Clin Dent 2015; 6: S86-89.
- 131) Schafer E, Olthoff G. Effect of three different sealers on the sealing ability of both thermafil obturators and cold laterally compacted gutta-percha. J Endod 2002; 28: 638-642.
- 132) Cobankara FK, Adanir N, Belli S, Pashley DH. A quantitative evaluation of apical leakage of four root-canal sealers. Int Endod J 2002; 35: 979-984.
- 133) Bouillaguet S, Shaw L, Barthelemy J, Krejci I, Wataha JC. Long-term sealing ability of Pulp Canal Sealer, AH-Plus, GuttaFlow and Epiphany. Int Endod J 2008; 41: 219-226.
- 134) Savariz A, Gonzalez-Rodriguez MP, Ferrer-Luque CM. Longterm sealing ability of GuttaFlow versus Ah Plus using different obturation techniques. Med Oral Patol Oral Cir Bucal 2010; 15: e936-941.
- 135) Nawal RR, Parande M, Sehgal R, Rao NR, Naik A. A comparative evaluation of 3 root canal filling systems. Oral Surg Oral Med Oral Pathol Oral Radiol Endod 2011; 111: 387-393.
- 136) Patni PM, Chandak M, Jain P, Patni MJ, Jain S, Mishra P,

et al. Stereomicroscopic evaluation of sealing ability of four different root canal sealers —an invitro Study. J Clin Diagn Res 2016; 10: ZC37-39.

- 137) Zmener O, Spielberg C, Lamberghini F, Rucci M. Sealing properties of a new epoxy resin-based root-canal sealer. Int Endod J 1997; 30: 332-334.
- 138) Siqueira JF Jr, Rocas IN, Valois CR. Apical sealing ability of five endodontic sealers. Aust Endod J 2001; 27: 33-35.
- 139) Miletic I, Ribaric SP, Karlovic Z, Jukic S, Bosnjak A, Anic I. Apical leakage of five root canal sealers after one year of storage. J Endod 2002; 28: 431-432.
- 140) Bodrumlu E, Avsar A, Meydan AD, Tuloglu N. Can radiotherapy affect the apical sealing ability of resin-based root canal sealers? J Am Dent Assoc 2009; 140: 326-330.
- 141) Bodrumlu E, Parlak E, Bodrumlu EH. The effect of irrigation solutions on the apical sealing ability in different root canal sealers. Braz Oral Res 2010; 24: 165-169.
- 142) Miletic I, Anic I, Pezelj-Ribaric S, Jukic S. Leakage of five root canal sealers. Int Endod J 1999; 32: 415-418.
- 143) Pappen AF, Bravo M, Gonzalez-Lopez S, Gonzalez-Rodriguez MP. An in vitro study of coronal leakage after intraradicular preparation of cast-dowel space. J Prosthet Dent 2005; 94: 214-218.
- 144) Cobankara FK, Orucoglu H, Sengun A, Belli S. The quantitative evaluation of apical sealing of four endodontic sealers. J Endod 2006; 32: 66-68.
- 145) Asawaworarit W, Yachor P, Kijsamanmith K, Vongsavan N. Comparison of the apical sealing ability of calcium silicatebased sealer and resin-based sealer using the fluid-filtration technique. Med Princ Pract 2016; 25: 561-565.
- 146) Ballullaya SV, Vinay V, Thumu J, Devalla S, Bollu IP, Balla S. Stereomicroscopic dye leakage measurement of six different root canal sealers. J Clin Diagn Res 2017; 11: ZC65-ZC68.
- 147) Altan H, Goztas Z, Inci G, Tosun G. Comparative evaluation of apical sealing ability of different root canal sealers. Eur Oral Res 2018; 52: 117-121.
- 148) De Almeida WA, Leonardo MR, Tanomaru Filho M, Silva LA. Evaluation of apical sealing of three endodontic sealers. Int Endod J 2000; 33: 25-27.
- 149) Biggs SG, Knowles KI, Ibarrola JL, Pashley DH. An in vitro assessment of the sealing ability of Resilon/Epiphany using fluid filtration. J Endod 2006; 32: 759-761.
- 150) Kaya BU, Kececi AD, Belli S. Evaluation of the sealing ability of gutta-percha and thermoplastic synthetic polymer-based systems along the root canals through the glucose penetration model. Oral Surg Oral Med Oral Pathol Oral Radiol Endod 2007; 104: e66-73.
- 151) Nagas E, Altundasar E, Serper A. The effect of master point taper on bond strength and apical sealing ability of different root canal sealers. Oral Surg Oral Med Oral Pathol Oral Radiol Endod 2009; 107: e61-64.
- 152) Onay EO, Ungor M, Orucoglu H. An in vitro evaluation of the apical sealing ability of a new resin-based root canal obturation system. J Endod 2006; 32: 976-978.
- 153) Tunga U, Bodrumlu E. Assessment of the sealing ability of a new root canal obturation material. J Endod 2006; 32: 876-878.
- 154) Bodrumlu E, Tunga U. Apical leakage of Resilon obturation material. J Contemp Dent Pract 2006; 7: 45-52.
- 155) Bodrumlu E, Tunga U. The apical sealing ability of a new root canal filling material. Am J Dent 2007; 20: 295-298.
- 156) Hirai VH, da Silva Neto UX, Westphalen VP, Perin CP, Carneiro E, Fariniuk LF. Comparative analysis of leakage in root canal fillings performed with gutta-percha and Resilon cones with AH Plus and Epiphany sealers. Oral Surg Oral Med Oral Pathol Oral Radiol Endod 2010; 109: e131-135.
- 157) Belli S, Ozcan E, Derinbay O, Eldeniz AU. A comparative evaluation of sealing ability of a new, self-etching, dualcurable sealer: hybrid root SEAL (MetaSEAL). Oral Surg

Oral Med Oral Pathol Oral Radiol Endod 2008; 106: e45-52.

- 158) Melih I, Jakovljevic A, Popovic M, Pesic D. [Comparative evaluation of sealing ability of different obturation materials]. Srp Arh Celok Lek 2010; 138: 287-291.
- 159) Fathia E, Hassan Abu-Bakr N, Yahia I. A comparative study of the microleakage of Resilon/Epiphany and gutta-percha/ AH-Plus obturating systems. Iran Endod J 2012; 7: 139-143.
- 160) Lambor RT, de Ataide Ide N, Chalakkal P, Akkara F, Shariff SA, Fernandes KS. An in vitro comparison between the apical sealing abilities of resilon with Epiphany sealer and guttapercha with AH plus sealer. Indian J Dent Res 2012; 23: 694.
- 161) Dhaded N, Dhaded S, Patil C, Patil R, Roshan JM. The effect of time of post space preparation on the seal and adaptation of resilon-Epiphany Se & gutta-percha-AH Plus sealer —an sem study. J Clin Diagn Res 2014; 8: 217-220.
- 162) Sultana M, Musani MA, Ahmed IM. An in-vitro comparative study for assessment of apical sealing ability of Epiphany/ AH Plus sealer with Resilon/gutta-percha root canal filling materials. J Int Soc Prev Community Dent 2016; 6: 321-326.
- 163) Kumar SA, Shivanna V, Naian MT, Shivamurthy G. Comparative evaluation of the apical sealing ability and adaptation to dentine of three resin-based sealers: An in vitro study. J Conserv Dent 2011; 14: 16-20.
- 164) Kardon BP, Kuttler S, Hardigan P, Dorn SO. An in vitro evaluation of the sealing ability of a new root-canal-obturation system. J Endod 2003; 29: 658-661.
- 165) Sevimay S, Kalayci A. Evaluation of apical sealing ability and adaptation to dentine of two resin-based sealers. J Oral Rehabil 2005; 32: 105-110.
- 166) Pawar SS, Pujar MA, Makandar SD. Evaluation of the apical sealing ability of bioceramic sealer, AH plus & Epiphany: An in vitro study. J Conserv Dent 2014; 17: 579-582.
- 167) Canadas PS, Berastegui E, Gaton-Hernandez P, Silva LA, Leite GA, Silva RS. Physicochemical properties and interfacial adaptation of root canal sealers. Braz Dent J 2014; 25: 435-441.
- 168) Ahuja L, Jasuja P, Verma KG, Juneja S, Mathur A, Walia R, et al. A comparative evaluation of sealing ability of new MTA based sealers with conventional resin based sealer: an in-vitro study. J Clin Diagn Res 2016; 10: ZC76-79.
- 169) Jain P, Pruthi V, Sikri VK. An ex vivo evaluation of the sealing ability of polydimethylsiloxane-based root canal sealers. Indian J Dent Res 2014; 25: 336-339.
- 170) Orstavik D, Nordahl I, Tibballs JE. Dimensional change following setting of root canal sealer materials. Dent Mater 2001; 17: 512-519.
- 171) Royer K, Liu XJ, Zhu Q, Malmstrom H, Ren YF. Apical and root canal space sealing abilities of resin and glass ionomerbased root canal obturation systems. Chin J Dent Res 2013; 16: 47-53.
- 172) Mejare I, Mjor IA. Glass ionomer and resin-based fissure sealants: a clinical study. Scand J Dent Res 1990; 98: 345-350.
- 173) Aptekar A, Ginnan K. Comparative analysis of microleakage and seal for 2 obturation materials: Resilon/Epiphany and gutta-percha. J Can Dent Assoc 2006; 72: 245.
- 174) Kumar RV, Shruthi C. Evaluation of the sealing ability of resin cement used as a root canal sealer: An in vitro study. J Conserv Dent 2012; 15: 274-277.
- 175) Imai Y, Komabayashi T. Properties of a new injectable type of root canal filling resin with adhesiveness to dentin. J Endod 2003; 29: 20-23.
- 176) Siqueira JF Jr, Rocas IN. Clinical implications and microbiology of bacterial persistence after treatment procedures. J Endod 2008; 34: 1291-1301 e1293.
- 177) Banoee M, Seif S, Nazari ZE, Jafari-Fesharaki P, Shahverdi HR, Moballegh A, *et al.* ZnO nanoparticles enhanced antibacterial activity of ciprofloxacin against Staphylococcus

aureus and Escherichia coli. J Biomed Mater Res B Appl Biomater 2010; 93: 557-561.

- 178) Poggio C, Lombardini M, Colombo M, Dagna A, Saino E, Arciola CR, *et al.* Antibacterial effects of six endodontic sealers. Int J Artif Organs 2011; 34: 908-913.
- 179) Heling I, Chandler NP. The antimicrobial effect within dentinal tubules of four root canal sealers. J Endod 1996; 22: 257-259.
- 180) Wainstein M, Morgental RD, Waltrick SB, Oliveira SD, Vier-Pelisser FV, Figueiredo JA, *et al.* In vitro antibacterial activity of a silicone-based endodontic sealer and two conventional sealers. Braz Oral Res 2016; 30: 1-5.
- 181) Kapralos V, Koutroulis A, Ørstavik D, Sunde PT, Rukke HV. Antibacterial activity of endodontic sealers against planktonic bacteria and bacteria in biofilms. J Endod 2018; 44: 149-154.
- 182) Torabinejad M, Hong CU, Pitt Ford TR, Kettering JD. Antibacterial effects of some root end filling materials. J Endod 1995; 21: 403-406.
- 183) Estrela C, Bammann LL, Estrela CR, Silva RS, Pecora JD. Antimicrobial and chemical study of MTA, Portland cement, calcium hydroxide paste, Sealapex and Dycal. Braz Dent J 2000; 11: 3-9.
- 184) Miyagak DC, de Carvalho EM, Robazza CR, Chavasco JK, Levorato GL. In vitro evaluation of the antimicrobial activity of endodontic sealers. Braz Oral Res 2006; 20: 303-306.
- 185) Tanomaru-Filho M, Tanomaru JM, Barros DB, Watanabe E, Ito IY. In vitro antimicrobial activity of endodontic sealers, MTA-based cements and Portland cement. J Oral Sci 2007; 49: 41-45.
- 186) Asgary S, Kamrani FA. Antibacterial effects of five different root canal sealing materials. J Oral Sci 2008; 50: 469-474.
- 187) Okiji T, Yoshiba K. Reparative dentinogenesis induced by mineral trioxide aggregate: a review from the biological and physicochemical points of view. Int J Dent 2009; 2009: 464280.
- 188) Singh G, Gupta I, Elshamy FM, Boreak N, Homeida HE. In vitro comparison of antibacterial properties of bioceramicbased sealer, resin-based sealer and zinc oxide eugenol based sealer and two mineral trioxide aggregates. Eur J Dent 2016; 10: 366-369.
- 189) Duarte MA, Demarchi AC, Yamashita JC, Kuga MC, Fraga Sde C. pH and calcium ion release of 2 root-end filling materials. Oral Surg Oral Med Oral Pathol Oral Radiol Endod 2003; 95: 345-347.
- 190) Shakya VK, Gupta P, Tikku AP, Pathak AK, Chandra A, Yadav RK, Bharti R, Singh RK. An in vitro evaluation of antimicrobial efficacy and flow characteristics for AH Plus, MTA Fillapex, CRCS and Gutta Flow 2 root canal sealer. J Clin Diagn Res 2016; 10: ZC104-108.
- 191) Zhang H, Shen Y, Ruse ND, Haapasalo M. Antibacterial activity of endodontic sealers by modified direct contact test against Enterococcus faecalis. J Endod 2009; 35: 1051-1055.
- 192) Tanomaru JM, Tanomaru-Filho M, Hotta J, Watanabe E, Ito IY. Antimicrobial activity of endodontic sealers based on calcium hydroxide and MTA. Acta Odontol Latinoam 2008; 21: 147-151.
- 193) Teoh YY, Athanassiadis B, Walsh LJ. Sealing ability of alkaline endodontic cements versus resin cements. Materials (Basel) 2017; 10: 1-7.
- 194) Gulati N, Chandra S, Aggarwal PK, Jaiswal JN, Singh M. Cytotoxicity of eugenol in sealer containing zinc-oxide. Endod Dent Traumatol 1991; 7: 181-185.
- 195) Kolokouris I, Economides N, Beltes P, Vlemmas I. In vivo comparison of the biocompatibility of two root canal sealers implanted into the subcutaneous connective tissue of rats. J Endod 1998; 24: 82-85.
- 196) Serene TP, Vesely J, Boackle RJ. Complement activation as a possible in vitro indication of the inflammatory potential of endodontic materials. Oral Surg Oral Med Oral Pathol 1988;

65: 354-357.

- 197) Guigand M, Pellen-Mussi P, Le Goff A, Vulcain JM, Bonnaure-Mallet M. Evaluation of the cytocompatibility of three endodontic materials. J Endod 1999; 25: 419-423.
- 198) Cintra LTA, Benetti F, de Azevedo Queiroz IO, Ferreira LL, Massunari L, Bueno CRE, et al. Evaluation of the cytotoxicity and biocompatibility of new resin epoxy-based endodontic sealer containing calcium hydroxide. J Endod 2017; 43: 2088-2092.
- 199) Pertot WJ, Camps J, Remusat M, Proust JP. In vivo comparison of the biocompatibility of two root canal sealers implanted into the mandibular bone of rabbits. Oral Surg Oral Med Oral Pathol 1992; 73: 613-620.
- 200) Nakamura H, Sakakibara F, Matsumoto Y, Hirano S, Hayakawa H, Sakai K, *et al.* Study on the cytotoxicity of root canal filling materials. J Endod 1986; 12: 156-160.
- 201) al-Khatib ZZ, Baum RH, Morse DR, Yesilsoy C, Bhambhani S, Furst ML. The antimicrobial effect of various endodontic sealers. Oral Surg Oral Med Oral Pathol 1990; 70: 784-790.
- 202) Dahl JE. Toxicity of endodontic filling materials. Endod Topics 2005; 12: 39-43.
- 203) Hume WR, Mount GJ. In vitro studies on the potential for pulpal cytotoxicity of glass-ionomer cements. J Dent Res 1988; 67: 915-918.
- 204) Zetterqvist L, Anneroth G, Nordenram A. Glass-ionomer cement as retrograde filling material. An experimental investigation in monkeys. Int J Oral Maxillofac Surg 1987; 16: 459-464.
- 205) Zetterqvist L, Anneroth G, Danin J, Roding K. Microleakage of retrograde fillings: a comparative investigation between amalgam and glass ionomer cement in vitro. Int Endod J 1988; 21: 1-8.
- 206) Briseno BM, Willershausen B. Root canal sealer cytotoxicity on human gingival fibroblasts: 2. Silicone- and resin-based sealers. J Endod 1991; 17: 537-540.
- 207) Silva EJ, Neves AA, De-Deus G, Accorsi-Mendonca T, Moraes AP, Valentim RM, *et al.* Cytotoxicity and gelatinolytic activity of a new silicon-based endodontic sealer. J Appl Biomater Funct Mater 2015; 13: e376-380.
- 208) Ko H, Yang W, Park K, Kim M. Cytotoxicity of mineral trioxide aggregate (MTA) and bone morphogenetic protein 2 (BMP-2) and response of rat pulp to MTA and BMP-2. Oral Surg Oral Med Oral Pathol Oral Radiol Endod 2010; 109: e103-108.
- 209) Benetti F, Queiroz ÍOdA, Cosme-Silva L, Conti LC, Oliveira SHPd, Cintra LTA. Cytotoxicity, biocompatibility and biomineralization of a new ready-for-use bioceramic repair material. Braz Dent J 2019; 30: 325-332.
- 210) Camps J, Jeanneau C, El Ayachi I, Laurent P, About I. Bioactivity of a calcium silicate-based endodontic cement (BioRoot RCS): interactions with human periodontal ligament cells in vitro. J Endod 2015; 41: 1469-1473.
- 211) Loushine BA, Bryan TE, Looney SW, Gillen BM, Loushine RJ, Weller RN, *et al.* Setting properties and cytotoxicity evaluation of a premixed bioceramic root canal sealer. J Endod 2011; 37: 673-677.
- 212) Huang FM, Tai KW, Chou MY, Chang YC. Cytotoxicity of resin-, zinc oxide-eugenol-, and calcium hydroxide-based root canal sealers on human periodontal ligament cells and permanent V79 cells. Int Endod J 2002; 35: 153-158.
- 213) Pascon EA, Leonardo MR, Safavi K, Langeland K. Tissue reaction to endodontic materials: methods, criteria, assessment, and observations. Oral Surg Oral Med Oral Pathol 1991; 72: 222-237.
- 214) Mutoh N, Satoh T, Watabe H, Tani-Ishii N. Evaluation of the biocompatibility of resin-based root canal sealers in rat periapical tissue. Dent Mater J 2013; 32: 413-419.
- 215) Hirabayashi C, Imai Y. Studies on MMA-TBB resin. I. Comparison of TBB and other initiators in the polymerization

of PMMA/MMA resin. Dent Mater J 2002; 21: 314-321.

- 216) Imai Y, Watanabe M, Lee HE, Kojima K, Kadoma Y. Cytotoxicity of monomers used in dental resins. Reports Inst Med Dent Eng 1988; 22: 87-90.
- 217) Takeda S, Hashimoto Y, Miura Y, Kimura Y, Nakamura M. Cytotoxicity test of dental monomers using serum-free cell culture (in vitro). Shika Zairyo Kikai 1993; 12: 613-619.
- 218) Yoshii E. Cytotoxic effects of acrylates and methacrylates: relationships of monomer structures and cytotoxicity. J Biomed Mater Res 1997; 37: 517-524.
- 219) Lodiene G, Kopperud HM, Orstavik D, Bruzell EM. Detection of leachables and cytotoxicity after exposure to methacrylateand epoxy-based root canal sealers in vitro. Eur J Oral Sci 2013; 121: 488-496.
- 220) Camargo CH, Oliveira TR, Silva GO, Rabelo SB, Valera MC, Cavalcanti BN. Setting time affects in vitro biological properties of root canal sealers. J Endod 2014; 40: 530-533.
- 221) Martinho FC, Camargo SEA, Fernandes AMM, Campos MS, Prado RF, Camargo CHR, *et al.* Comparison of cytotoxicity, genotoxicity and immunological inflammatory biomarker activity of several endodontic sealers against immortalized human pulp cells. Int Endod J 2018; 51: 41-57.
- 222) Garza EG, Wadajkar A, Ahn C, Zhu Q, Opperman LA, Bellinger LL, *et al.* Cytotoxicity evaluation of methacrylate-

based resins for clinical endodontics in vitro. J Oral Sci 2012; 54: 213-217.

- 223) Leonardo MR, Silva LA, Utrilla LS, Assed S, Ether SS. Calcium hydroxide root canal sealers--histopathologic evaluation of apical and periapical repair after endodontic treatment. J Endod 1997; 23: 428-432.
- 224) Poggio C, Riva P, Chiesa M, Colombo M, Pietrocola G. Comparative cytotoxicity evaluation of eight root canal sealers. J Clin Exp Dent 2017; 9: e574-e578.
- 225) Kakehashi S, Stanley HR, Fitzgerald RJ. The effects of surgical exposures of dental pulps in germ-free and conventional laboratory rats. Oral Surg Oral Med Oral Pathol 1965; 20: 340-349.
- 226) Madison S, Swanson K, Chiles SA. An evaluation of coronal microleakage in endodontically treated teeth. Part II. Sealer types. J Endod 1987; 13: 109-112.
- 227) Ray HA, Trope M. Periapical status of endodontically treated teeth in relation to the technical quality of the root filling and the coronal restoration. Int Endod J 1995; 28: 12-18.
- 228) Gandolfi MG, Parrilli AP, Fini M, Prati C, Dummer PM. 3D micro-CT analysis of the interface voids associated with Thermafil root fillings used with AH Plus or a flowable MTA sealer. Int Endod J 2013; 46: 253-263.